Emergence of multi-drug resistant Non-fermenting gramnegative bacteria in causing Hospital-acquired infections in ICU setting of a tertiary care hospital.

- Das Gupta, Subhayan; MD Microbiology, Malda Medical College and Hospital. 1.
- 2. Das Roy, Rohon; MD Microbiology, Midnapore Medical College and Hospital.
 - 1. Department of Microbiology, Malda Medical College and Hospital.
 - 2. Department of Microbiology, Midnapore Medical College and Hospital.
 - Corresponding Author: Dr. Rohon Das Roy.

Abstract

Aims: This study evaluated the emergence of multidrug resistant non-fermenting bacteria causing nosocomial infections in intensive care units.

Methods and Results: A prospective study was carried out in patients who were admitted to ICU and underwent ventilation, catheterization, had a central-line insertion, or developed wound infection and then started showing features of invasive sepsis 48 hours after their admission, were included. Endotracheal aspirate/Bronchoalveolar lavage, urine, central-line catheters and pus samples were collected for culture and antibiotic sensitivity. 126 out of 224 samples showed growth on culture media. VAP accounted for 42% of all nosocomial infections. Acinetobacter baumannii from respiratory and blood samples were susceptible only to Polymyxin B and doxycycline to some extent. Pseudomonas aeruginosa from respiratory and urine samples was sensitive to Polymyxin B.

Conclusion: Emerging multi-drug resistance in Acinetobacter baumannii is becoming a difficult problem to tackle with.

Significance and Impact of Study: With the advent of new drugs, a concurrent rise in multi-drug resistant organisms has been seen with high virulent properties that is severely affecting the mortality rates. Nonfermenters, especially Acinetobacter baumanniiand Pseudomonas aeruginosa are the two most frequently isolated organisms among the bacterial pathogens which have developed resistance to most of the anti-bacterial agents.

Keywords: Acinetobacter baumannii; multi-drug resistant; nosocomial infections; Polymyxin B; Pseudomonas aeruginosa

Date of Submission: 06-08-2021

Date of Acceptance: 19-08-2021 _____

Introduction I.

Hospital Acquired Infections (HAI) or nosocomial infections are defined as those infections that are not present or incubating at the time of admission and manifest 48 hours after admission to the hospital.¹ HAIs are the leading cause of morbidity and mortality among hospitalized patients.

Healthcare associated infections in Intensive Care Units (ICU) in India range from 9.6% to 17.7% ^{2,3} The risk of HAI in ICU is 5 to 10 times greater than those acquired in general medical and surgical wards.A probable explanation for this may be the frequent requirement of invasive medical devices such as urinary catheters, central venous and arterial catheters and endotracheal tubes by critically ill patients, thus compromising normal skin and mucosal barriers.⁴

Among the nosocomial infections, ventilator-associated pneumonia (VAP), catheter-associated urinary tract infection (CAUTI) and central line-associated bloodstream infections (CRBSI) are seen in approximately 30% of ICU patients. The pathogens responsible for a substantial proportion of HAIs include vancomycinresistant enterococci (VRE), methicillin-resistant Staphylococcus aureus (MRSA), Klebsiella pneumoniae that produce extended-spectrum beta-lactamases, carbapenem-resistant Acinetobacter baumannii and Pseudomonas aeruginosaand Enterobacter spp. Other pathogens, such as, Escherichia coli, Candida albicans, Clostridium difficile, etc also play vital role in the occurrence of nosocomial infections. 5,6,7

Antibiotic resistance among bacterial pathogens isolated in ICU infections has become a major problem worldwide. Treatment becomes more challenging in case of gram-negative organisms causing lifethreatening infections in ICU, such as pneumonia, bloodstream infections, surgical site infections and meningitis; mainly due to the increasing multidrug resistance rates and limited novel antimicrobial agents.

It is evident that these nosocomial infections in ICUs play a vital role in the increased duration of hospital stay, cost of treatment, higher morbidity and mortality rates. Hence, this study was done to determine the prevalence and nature of nosocomial infections in the ICU of a government tertiary care hospital in West Bengal, India, evaluate the bacteriological profile of the isolates and analyse the drug sensitivity pattern of the isolated pathogen.

II. Methods

Study Design and Setting:

A prospective analytical study was carried out in the Department of Microbiology in collaboration with the Department of Anaesthesiology and Critical Care Medicine & Department of Surgery in a tertiary care government medical college in West Bengal, India.

Study Population:

The study only included patients who were intubated and on mechanical ventilation, patients with urinary catheters, with central venous catheters or with wound infections and started developing clinical features of invasive sepsis (systemic inflammatory response syndrome, SIRS) 48 hours after admission in the ICU. Their samples were collected for bacteriological culture and antibiotic susceptibility.Patients admitted to the ICU with sepsis were not included unless they developed new signs of SIRS more than 48 hours after admission. Patients having infections prior to admission in ICU and those admitted in ICU without infections were also excluded from the study.

Sample processing:

Blood cultures were processed using the BACT/ALERT 3D system (BioMerieux, Marcy L'Etoile, France) and incubated at 36°C for 7 days. When the culture bottle flagged positive, gram stain and subculture was performed. The respiratory samples (endotracheal aspirate, bronchoalveolar lavage), pus, urine and central venous catheters were plated on blood agar and MacConkey agar. All plates were incubated overnight at 37°C.A colony count of >10⁵ Colony forming units(CFU)/ mL was considered significant for endotracheal aspirate and urine samples. For BAL fluid, a colony count of 10^4 bacterial colonies per ml was considered significant.^{9,10}For central venous catheter, significant growth was defined as \geq 15 colonies by roll plate Method and \geq 100 CFU/ml by flush method.¹¹ Antimicrobial susceptibility of the isolates were tested on Mueller-Hinton agar by modified Kirby Bauer disk diffusion method as per the recommendations of Clinical and Laboratory Standards Institute (CLSI)-2020. Vancomycin E-test was used to determine its susceptibility in MRSA isolates.

III. Results

In this study, a total of 224 samples from patients admitted in ICU were processed in the laboratory, out of which only, 126 (56.25%) of the samples were culture positive. 86% of respiratory samples, 38.01% of urine samples, 100% of pus samples and 40% of blood samples showed culture positivity. [Table 1]

Type of sample	Culture positive	Culture positive	Culture negative	Culture negative		
	No.	%	No.	%		
Respiratory samples(n=64)	55	86	9	14		
Urine (n=54)	21	38.01	33	61.99		
Pus (n=12)	12	100	0	0		
Blood (n=94)	38	40	56	60		

TABLE 1: Correlation between samples and culture positivity.

Pneumonia accounted for 42% of all the nosocomial infections, followed by blood stream infections (33.5%) and urinary tract infections. (17.7%). *Acinetobacter baumannii* was found to be the most common bacteria causing blood stream infections (43%) and pneumonia (58%). Urinary Tract Infection and wound infections among ICU patients were mostly caused by *Escherichia coli* (54.5% and 48% respectively). [Table 2]

	Pneumonia N=53	Urinary Tract Infection N=22	Wound infection N=9	Blood Stream Infection N=42
Acinetobacter baumanii	58% N=31	_	_	43% N=18
Escherichia coli	5%	54.5%	48%	7%
	N=3	N=12	N=4	N=3
Klebsiella pneumoniae	11%	18.3%	18.7%	23%
pneumoniue	N=6	N=4	N=2	N=10
Staphylococcus aureus	3.8%			10
	N=2	-	-	N=4
Enterococcus spp.	3.8% N=2	-	-	7% N=3
Staphylococcus epidermidis	-	-	-	10% N=4
Pseudomonas aeruginosa	14% N=7	13.6% N=3	-	-
Proteus mirabilis		13.6%		
	-	N=3	-	-
Enterobacter spp.	3.8% N=2	-	33.3% N=3	-

Table 2: List of organisms isolated causing nosocomial infections in ICU.

Acinetobacter baumannii isolates from both blood and respiratory samples showed a good sensitivity to Polymyxin B (88.9% and 80.6% respectively) and was moderately sensitive to Doxycycline (50% and 41.9% respectively). Meropenem had no sensitivity against the isolates from blood, and around 10% susceptibility toisolates from respiratory samples. *Enterobacteriaceae* isolates from blood, respiratory samples and pus showed high level resistance to Meropenem, whereas 75% of the isolates from urine were sensitive to Meropenem. Sensitivity was practically absent to Ciprofloxacin. *Pseudomonas aeruginosa* isolated from respiratory samples and urine showed sensitivity mostly to Polymyxin B (71.4% and 66.7% respectively) and Aztreonam (42.8% and 66.7% respectively), while being resistant to most other antibiotics. [Table 3]

HAI	Causative organism	PIT	CIP	MRP	PB	3 rd GEN CEP	DOX	СОТ	AZM	GEN	NIT
Pneumoni a	Acinetobact er baumanii	0%	0%	10%	80.6 %	10%	41.9 %	12.7 %	25.8 %	25.8 %	-
	Escherichia coli	33.3 %	33.3 %	33.3 %	100%	33.3 %	33.3 %	33.3 %	33.3 %	33.3 %	-
	Klebsiella pneumoniae	16.7 %	16.7 %	16.7 %	83.3 %	16.7 %	33.3 %	16.7 %	33.3 %	33.3 %	-
	Pseudomona s aeruginosa	0%	0%	28.5 %	71.4 %	0%	0%	-	42.8 %	28.5 %	-
	Enterobacte r spp.	50%	50%	50%	100%	50%	50%	50%	50%	50%	-
Urinary Tract Infection	Escherichia coli	33.3 %	0%	75%	75%	33.3 %	_	-	50%	75%	33.3 %
	Klebsiella pneumoniae	25%	0%	75%	75%	25%	-	-	50%	75%	25%
	Pseudomona s aeruginosa	33.3 %	0%	33.3 %	66.7 %	33.3 %	-	-	66.7 %	66.7 %	66.7 %
	Proteus mirabilis	33.3 %	0%	66.7 %	0%	33.3 %	-	-	66.7 %	66.7 %	33.3 %
Wound infection	Escherichia coli	25%	0%	25%	75%	0%	25%	25%	25%	25%	-
	Klebsiella pneumoniae	0%	0%	50%	100%	0%	50%	0%	0%	0%	-
	Enterobacte r spp.	33.3 %	0%	33.3 %	66.7 %	0%	33.3 %	33.3 %	33.3 %	33.3 %	-

Emergence of multi-drug resistant Non-fermenting gram-negative bacteria in causing..

Blood	Acinetobact	0%	0%	0%	88.9	11.1	50%	34%	27.7	22.3	
Stream	er baumanii				%	%			%	%	
Infection											-
	Escherichia	33.3	33.3	33.3	100%	33.3	66.7	33.3	66.7	33.3	
	coli	%	%	%		%	%	%	%	%	
											-
	Klebsiella	40%	10%	10%	100%	40%	30%	10%	20%	40%	
	pneumoniae										
											-

Table 4: Antibiotic sensitivity rates of different gram-negative isolates.

[PIT: Piperacillin-Tazobactam, CIP: Ciprofloxacin, MRP: Meropenem, PB: Polymyxin B, CEP: Cephalosporin; Ceftazidime was used for *Acinetobacter baumannii* and *Pseudomonas aeruginosa*; Ceftriaxone was used for Enterobacteriaceae. DOX: Doxycycline, COT: Cotrimoxazole, AZM: Azithromycin NIT: Nitrofurantoin]

All gram-positive isolates from respiratory samples were 100% sensitive to Vancomycin and Linezolid, however, two Linezolid resistant *Staphylococcus epidermidis* (LRSE) isolates were found in blood samples. All *Staphylococcus aureus* isolates were Cefoxitin resistant strains (Methicillin resistant *Staphylococcus aureus*). [Table 4]

		VA	LZ	CX	CXM	CD	RO	GEN	CIP
Pneumonia	Staphylococcus aureus	100%	100%	0%	0%	0%	0%	100%	50%
	Enterococcus spp.	100%	100%	-	-	0%	0%	100%	0%
Blood Stream Infection	Staphylococcus aureus	100%	100%	0%	0%	50%	50%	25%	0%
	Enterococcus spp.	100%	100%	-	-	_	66.7%	66.7%	0%
	Staphylococcus epidermidis	100%	50%	-	25%	50%	50%	25%	0%

Table 5: Antibiotic sensitivity rates of different gram-positive isolates.

[VA: Vancomycin, LZ: Linezolid, CX: Cefoxitin, CXM: Cefuroxime, CD: Clindamycin, RO: Roxithromycin, GEN: Gentamicin, CIP: Ciprofloxacin.]

IV. Discussion

Nosocomial infections in intensive care units with increasing incidence of multidrug resistance organisms is a matter of serious concern today. A proper understanding of the source of infection, the pathogens

involved and administration of drugs which have proven to be susceptible to these resistant organisms can significantly help to decrease the morbidity and mortality associated with these hospital-acquired infections. This study was focussed on understanding theemerging drug resistant pathogens causing different types of HAIs in ICU patients. Out of all the infections reported, ventilator associated pneumonia had the highest incidence rate of 42% followed by central-line blood stream infections (33.5%) and urinary tract infections (17.7%). European one-day prevalence study (EPIC)¹² conducted in 1992 showed a similar distribution of ICU-acquired infections with 46.9% cases of pneumonia, 17.6% urinary tract infections and 12% bloodstream infections. The study by Richards M J et al.¹³ reported a 31% occurrence of nosocomial pneumonia, 23% urinary tract infections and 14% primary bloodstream infections out of all nosocomial infections in a surgical ICU.

Acinetobacter baumannii was found to be the commonest organism causing blood stream infections (43%) and pneumonia (58%) in this study. In the study by Chopra T etal.¹⁴*Acinetobacter baumannii* was observed as themain organism causing bloodstream infections in health care settings.Hartzell JD etal.¹⁵studied on the increasing incidence of multidrug resistant strains of *Acinetobacter baumannii* causing nosocomial pneumonia.However, in another study by Kombadeet al.¹⁶from India in 2010, *Pseudomonas aeruginosa* (23.8%) was the commonest isolate obtained in respiratory samples, followed by *Acinetobacter baumannii*(21.2%) and *Klebsiellapneumoniae* (15.2%).

In the present study, Urinary Tract Infection was mostly caused by *Escherichia coli* (54.5%), followed by *Klebsiella pneumoniae* (18.3%), *Pseudomonas aeruginosa* and *Proteus mirabilis* both accounting 13.6%. Vyawahare CR et al.²¹ in his study on the occurrence of catheter-associated urinary tract infection in critical care units found *E. coli* 39 (57%) as the most common bacterial pathogen, followed by *Klebsiella spp.* 14 (20%), and *Pseudomonas spp.* 5 (7%). Infact, Karina et al.²² found a similar pattern in his study with *E. coli*, being the most common urinary pathogen (27%).

All gram-positive isolates from respiratory samples were sensitive to Vancomycin and Linezolid, however, two Linezolid resistant *Staphylococcus epidermidis* (LRSE) isolates were found in blood samples. A similar finding was observed in a study by Stephanie A Folan et al.¹⁷ where 40% of *Staphylococcus epidermidis* isolates causing bloodstream infections were Linezolid resistant.

Acinetobacter baumannii from both blood and respiratory samples showed 88.9% and 80.6% sensitivity to Polymyxin B respectively and was moderately sensitive to Doxycycline. Ciprofloxacin was found to be the least susceptible drug in this study. The study by Ngamprasertchai T et al.¹⁸ also observed similar results in pneumonia patientswhere 51.5% multidrug resistant *Acinetobacter spp.* isolates were susceptible to colistin.In the 2009 MYSTIC report,¹⁹ 67% of *Acinetobacter* isolates were resistant to ciprofloxacin.

Pseudomonas aeruginosa isolated from respiratory samples and urine showed 71.4% and 66.7% susceptibility to Polymyxin B respectively and also moderately to Aztreonam (42.8% and 66.7% respectively). *Pseudomonas aeruginosa* causing UTI showed 96% susceptibility to colistin and only 54% to aztreonam in the study done by Lamas Ferreiro JL et al.²⁰

V. Conclusion

Increasing incidence of *Acinetobacter baumannii* causing nosocomial pneumonia and blood stream infections and resistant to almost all antibiotics but Polymyxin B is a matter of grave importance.Carbapenems like meropenem and fluoroquinolones which used to have very good activity against gram negative pathogens, are almost of no use nowadays. Doxycycline still has a moderate susceptibility to these multidrug resistant pathogens. Linezolid still remains to be a very active drug against gram positive infections, though LRSE strains among central-line associated blood stream infections are frequently documented these days.

Author Contributions

Both listed authors have agreed to be the authors for this research paper and have contributed equally to the work's conception, design and performed the acquisition of data for the work.

Financial disclosure: The author(s) received no specific funding for this work.

Conflicts of Interest: There are no financial or personal conflicts of interest.

Reference

- Boev C, Kiss E. Hospital Acquired Infections: Current Trends and Prevention. Crit Care Nurs Clin North Am. 2017 Mar;29(1):51-65.
- [2]. Pradhan NP, Bhat SM, Ghadage DP. Nosocomial infections in the medical ICU: a retrospective study highlighting their prevalence, microbiological profile and impact on ICU stay and mortality. J Assoc Physicians India. 2014;62(10):18–21.
- [3]. Mythri H, Kashinath K. Nosocomial infections in patients admitted in intensive care unit of a tertiary health center, India. Ann Med Health Sci Res. 2014;4(5):738–741. doi: 10.4103/2141-9248.141540.
- [4]. Singh C, Chaturvedi R, Garg B, Datta R, Kumar A. Incidence of healthcare associated infection in the surgical ICU of a tertiary care hospital. Medical Journal Armed Forces India 69 2013; 124-129
- [5]. Vincent JL, Rello J, Marshall J, Silva E, Anzueto A, Martin CD. International study of the prevalence and outcomes of infection in intensive care units. JAMA 2009 Dec;302(21):2323-9.

- [6]. National Nosocomial Infections Surveillance (NNIS) system report, data summary from January 1992-April 2000, issued June 2000. Am J Infect Control 2000; 28:429
- [7]. Hidron AI, Edwards JR, Patel J, et al. NHSN annual update: antimicrobial-resistant pathogens associated with healthcare-associated infections: annual summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006-2007. Infect Control Hosp Epidemiol 2008; 29:996.
- [8]. Rahal JJ. Antimicrobial resistance among and therapeutic options against gram-negative pathogens. Clin Infect Dis 2009 Aug;49(Suppl 1): S4-10
- Betty A.Forbes, Daniel F Sahm, Alice S.Weissfeld: pirainfections of the Lower Respiratory Tract, In Bailey & Scott's Diagnostic Microbiology.12thed.Missouri, Mosby Elsevier,2007. P798-813.
- [10]. Jakribettu R, Boloor R.Characterisation of aerobic bacteria isolated from endotracheal aspirate in adult patients suspected ventilator associated pneumonia in a tertiary care center in Mangalore. Saudi J Anaesth. 2012 Apr-Jun; 6(2): 115–119.
- [11]. Parameswaran R, Sherchan JB, Varma DM, Mukhopadhyay C, Vidyasagar S.Intravascular Catheter related infection in a tertiary care hospital.J Infect Dev Ctries 2011; 5(6):452-458.
- [12]. Vincent J, Bihari DJ, Suter PM, Bruining HA, White J, Nicolas-Chanoin M, Wolff M, Spencer RC & Hemmer M (1995) The Prevalence of Nosocomial Infection in Intensive Care Units in Europe: Results of the European Prevalence of Infection In Intensive Care (EPIC) Study. JAMA 274(8): 639-644.
- [13]. Richards MJ, Edwards JR, Culver DH, Gaynes RP. Nosocomial infections in combined medical-surgical intensive care units in the United States. Infect Control Hosp Epidemiol. 2000 Aug;21(8):510-5. doi: 10.1086/501795. PMID: 10968716.
- [14]. Chopra T, Marchaim D, Awali RA, et al. Epidemiology of bloodstream infections caused by Acinetobacter baumannii and impact of drug resistance to both carbapenems and ampicillin-sulbactam on clinical outcomes. Antimicrob Agents Chemother. 2013;57(12):6270-6275. doi:10.1128/AAC.01520-13.
- [15]. Hartzell JD, Kim AS, Kortepeter MG, Moran KA. Acinetobacter pneumonia: a review. MedGenMed. 2007;9(3):4. Published 2007 Jul 5.
- [16]. S.P.Kombade1 and G.N.Agrawal. Microbiological study of lower respiratory tract infections in ICU patients. *Int.J.Curr.Microbiol.App.Sci*(2014) 3(8) 749-754
- [17]. Stephanie A Folan, Kayleigh R Marx, Frank P Tverdek, IssamRaad, Victor E Mulanovich, Jeffrey J Tarrand, Samuel A Shelburne, Samuel L Aitken, Clinical Outcomes Associated With Linezolid Resistance in Leukemia Patients With Linezolid-Resistant Staphylococcus epidermidis Bacteremia, Open Forum Infectious Diseases, Volume 5, Issue 7, July 2018, ofy167, https://doi.org/10.1093/ofid/ofy167
- [18]. Ngamprasertchai T, Boonyasiri A, Charoenpong L, et al. Effectiveness and safety of polymyxin B for the treatment of infections caused by extensively drug-resistant Gram-negative bacteria in Thailand. *Infect Drug Resist.* 2018;11:1219-1224. Published 2018 Aug 20. doi:10.2147/IDR.S169939.
- [19]. Rhomberg PR, Jones RN. Summary trends for the Meropenem Yearly Susceptibility Test Information Collection Program: a 10year experience in the United States (1999-2008). Diagn Microbiol Infect Dis 2009; 65:414.
- [20]. Lamas Ferreiro JL, Álvarez Otero J, González González L, et al. Pseudomonas aeruginosa urinary tract infections in hospitalized patients: Mortality and prognostic factors. *PLoS One*. 2017;12(5):e0178178. Published 2017 May 26. doi:10.1371/journal.pone.0178178.
- [21]. Vyawahare CR, Gandham NR, Misra RN, Jadhav SV, Gupta NS, Angadi KM. Occurrence of catheter-associated urinary tract infection in critical care units. Med J DY Patil Univ2015;8:585-9.
- [22]. Karina BD, Myrna MT, Tessa TT. Catheter related Urinary tract infections: Incidence, risk factors and microbiologic profile. Phil J Microbiol Infect Dis 1999;28:133-8.

Dr. Rohon Das Roy, et. al. "Emergence of multi-drug resistant Non-fermenting gram-negative bacteria in causing Hospital-acquired infections in ICU setting of a tertiary care hospital." *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS)*, 20(08), 2021, pp. 17-20.
