

Prevalence and Antimicrobial Susceptibility pattern of Methicillin Resistant Staphylococcus aureus in Different Clinical Infections

Dr. Prasana Gupta¹, Mr Ravi Vashisth², Dr. Atul Kumar³, Mrs Malini Sharma⁴

¹HOD & Professor in Microbiology Department At NIMS University, Jaipur, Rajasthan, India

²Tutor in Microbiology Department At Muzaffarnagar Medical College, Muzaffarnagar, UP, India

³Assistant Professor In Microbiology Department At Muzaffarnagar Medical College, Muzaffarnagar, UP, India

Corresponding Author: MR. RAVI VASHISTHA

Abstract: Staphylococcus aureus is also known as "golden staph" and Oro staphira, it is facultative anaerobic Gram-positive cocci. It is frequently found as part of the normal skin flora on the skin and nasal passages. S. aureus is the most common species of Staphylococcus to cause Staph infections. Total 171 samples were taken and staphylococcus aureus were isolates from clinical samples. Microscopic observation of smears Gram positive cocci seen in clusters were observed. Observation of growth of clinical samples on the following medium Blood agar, MacConkey agar. In Coagulase test Positive slide coagulase & Tube coagulase tests. Urease Positive. Mannitol Ferments mannitol. Confirmed isolates of Staph aureus were subjected to following test. Among all 171 staphylococcus aureus MRSA and MSSA were also isolates. MRSA were 31(17%) MSSA 140(81%). As compare to sex distribution female 10(20%) were more infected than male 21(80%). Prevalence of MSSA age wise was 71-80 years = 100%, 1-10 years is 80%, 21-30 years is 77%. Prevalence of MRSA age wise was 41-50 years. is 22%, 11-20 years is 21%, 31-40 years is 20%.

Keywords: screening of Methicillin Resistant S.aureus by disc diffusion method.

Date of Submission: 26-01-2019

Date of acceptance: 09-02-2019

I. Introduction

Staphylococcus aureus very commonly causes infections in humans, virtually every person will have one or more Staphylococcus aureus infections in his or her lifetime¹. Staph .aureus ranks second as the cause of nosocomial blood stream infection that leads to increased morbidity, mortality, hospital stay, and costs². Nasal carriers of Staphylococcus aureus are also at increased risk of developing Staph. Aureus infection³. It is also one of the most common human pathogens, which cause a broad spectrum of illnesses ranging from relatively mild skin infection to life threatening septicemia⁴. As compare to other major human pathogens, antimicrobial resistance of staphylococcus aureus emerged along with the discovery and wide spread use of different classes of antibiotics.⁵. By contrast the VISA Prevalence rate was generally lower than 1% across the globe.⁶⁻⁹. Staph .aureus is an important human pathogen frequently isolated from systemic infections, community acquired & nosocomial infections. It is the part of normal human microflora, found in external environment, anterior nares, intertriginous skin folds, perineum, axilla, & vagina. Penicillin was the drug of choice for treatment. Resistance to penicillin Beta-lactamase. Infection caused by Staph .aureus, especially MRSA are emerging as a major public health problem in hospitals and community. The emergence and spread of both health care and community associated MRSA has made infection control intervention and treatment challenging¹. S. aureus produce numerous virulence factor including Panton Valentine Leucocidin {PVL} which is a pore forming cytotoxic more often identified community associated MRSA strain than hospital associated strains¹⁰. Isolation of VRSA from different countries has confirmed that the emergence of these strains is a globe issue¹¹⁻¹⁴

II. Methods And Materials

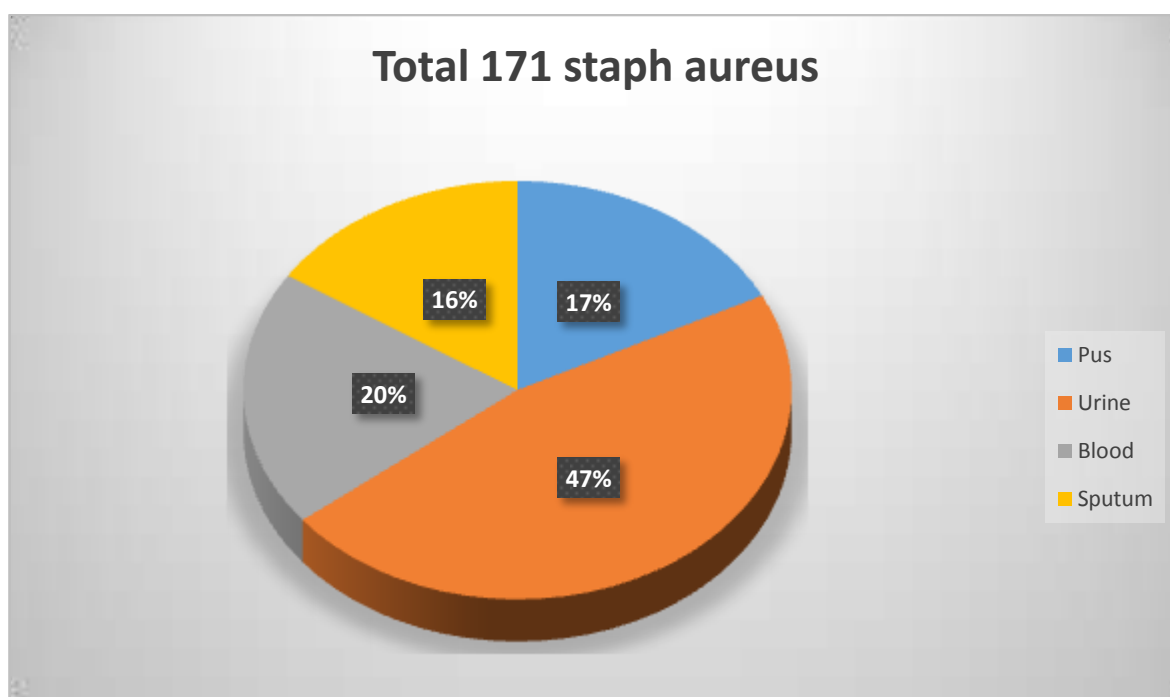
This study was conducted at Muzaffarnagar medical college and hospital, Muzaffarnagar for the period of one year (July 2017-july 2018). Various isolates of Staph aureus were obtained from different clinical samples Blood, Pus, Sputum, Endotracheal secretion, Catheter tips, Urine, CSF, Various body fluids (pleural, ascetic, synovial etc.) Microscopic observation of smears Gram positive cocci seen in clusters were observed. Observation of growth of clinical samples on the following medium Blood agar: Beta-hemolytic yellow orange colonies were seen. In MacConkey agar Small lactose fermenting colonies were seen. Secondary smears were made & observed before going on to identification steps. Identification of Staph aureus by Catalase Prompt positive reaction. In Coagulase test Positive slide coagulase & Tube coagulase tests. Urease Positive. Mannitol Ferments mannitol. Confirmed isolates of Staph aureus were subjected to following test Antibiotic sensitivity testing by following Kirby-Bauer Method/Disc Diffusion method using discs for gram positive Cocci. Screening

Prevalence and Antimicrobial Susceptibility pattern of Methicillin Resistant Staphylococcus aureus in .

for MRSA: MHA with 4% salt concentration were inoculated & filter paper disc containing 1µg oxacillin, were placed on it. Incubated at 35°C for 24 hrs. The zone of inhibition were studied and compared with standard chart. Susceptible (13mm), Intermediate (11-12mm), Resistant (10mm). In Screening for VISA: MHA plates were inoculated & filter paper disc containing 30µg vancomycin, were placed on it then Incubated at 35°C for 24 hrs. The zone of inhibition were studied and compared with standard chart. Susceptible (15mm) Intermediate (11-14mm) Resistant (10mm) Antibiotic resistance of the strains with other antibiotics Sensitivity pattern of MRSA & MSSA strains reference to other available antibiotics used for Staph aureus, were studied & compared. Antibiotic resistance of the strains with other antibiotics. Sensitivity pattern of MRSA & MSSA strains in reference to other available antibiotics used for Staph aureus, were studied & compared by CLSI (2009) guidelines¹⁵. Antibiotic sensitivity testing by following Kirby-Bauer Method/Disc Diffusion method Ampicillin/Sulbactam 20 mcg, Tetracycline 25 mcg, Cefotaxime 30 mcg, Ciprofloxacin 30 mcg, Levofloxacin 30 mcg, Gentamicin 5 mcg, Co-trim oxazole 5 mcg, Cephalexin 30mcg, Linezolid 1 mcg, Roxythromycin 2 mcg, Leucomycin 10 mcg.

III. Result

During the study period total 171 samples were taken and staphylococcus aureus were isolates from clinical samples such as Pus (30), Urine (80), Blood (34), and Sputum (27) were isolated.



Among all 171 staphylococcus aureus MRSA, MSSA were also isolates. MRSA were 31(17%) MSSA 140(81%) As compare to sex distribution female 10(20%) were more infected than male 21(80%). Prevalence of MRSA, MSSA in OPD and IPD were shown in figure1. Prevalence of MSSA age wise was 71-80 years = 100%, 1-10 years is 80%, 21-30 years is 77%.Prevalence of MRSA age wise was 41-50 years. is 22%, 11-20 years is 21%,31-40 years is 20%. Resistance of MRSA strains Ciprofloxacin 45% > Roxythromycin 43% > Cefotaxime (26%). Sensitivity of MRSA strains Oxacillin 90% > Ampicillin 87% > Linezolid & Levofloxacin 87%. Resistance of MSSA strain Ciprofloxacin 15% > Tetracycline 9% > Cotrimoxazole & Cefotaxime 8%. Sensitivity of MSSA strains: Linezolid 93% > Leucomycin & Ampicillin (94% > Oxacillin, Gentamicin & Cephalexin 92%.

Prevalence of staph aureus	IPD	OPD
MRSA	11	20
MSSA	97	43

IV. Conclusion & Discussion

MRSA is now a problem in hospitals worldwide and is being increasingly recovered from nursing homes and the community. In our study the prevalence of MRSA is 17%. Similar prevalence was shown by shilpi rao et al is 46 %¹⁶. Other studies have also shown such a high MRSA prevalence in various parts of the country ranging

Prevalence and Antimicrobial Susceptibility pattern of Methicillin Resistant Staphylococcus aureus in .

from 40.6% to 54.85% to 59.3%.^{17,18,19} However, 31.1 and 23.6% MRSA prevalence has also been reported²⁰⁻²¹. In Maulana Azad Medical College, New Delhi the prevalence of MRSA has increased from 9.83% (1988) to 45.44% (2009). Infection caused by drug resistance staphylococcus aureus are increasing worldwide due to emergence of MRSA. The prompt and accurate detection of MRSA isolate could be useful for determining the appropriate treatment strategy and subsequent effective management and control of corresponding infections. We found that the Prevalence of MRSA was 17% which correlates with previous reports. From other regions of countries Bangladesh, and Pakistan²²⁻²⁵. In my study we can conclude the following. The prevalence of MRSA is 31(17%). Maximum numbers of MRSA were isolated from urine samples. Most of the isolates of MRSA were resistant to ciprofloxacin, Roxythromycin, & tetracycline. Further studies are required to define setting specific strategies to reduce morbidity & mortality from S. aureus infections, prevention of MRSA transmission and emergence of drug resistance throughout the developing world.

References

- [1]. Okunye Olufemi L, Adeleke Olufemi E, Adegoke O Andrew and Adeyemi Omokehinde H. Antibiotic Susceptibility Pattern and Beta-lactamase Production in Isolates of Staphylococcus aureus from Recurrent Furunculosis in Southwestern, Nigeria. Sierra Leone Journal of Biomedical Research ISSN 2076-6270 (Print) Vol. 3(3) pp. 123-127, December, 2011.
- [2]. Jamshaid ali khan, Zafar iqbal, Liaqat ali and Abbas khan. Prevalence and β -lactamase producing bacterial isolates in a teaching hospital in Peshawar, Pakistan: a four year study Department of Pharmacy University of Peshawar, Peshawar, Pakistan. Postgraduate Medical Institute, Hayatabad Medical Complex, Peshawar
- [3]. Christoph K. Naber. Staphylococcus aureus Bacteremia: Epidemiology, Pathophysiology, and Management Strategies, Department of Cardiology, West-German Heart Center, Essen, Germany
- [4]. Lowy F.D staphylococcus aureus infections N Engl J Med 1988; 339:520-32
- [5]. Lowy F.D Antimicrobial resistance; the example of D staphylococcus aureus. J clin invest 2003; 111:1265-73
- [6]. HO CM, Hsueh PR, liu CY, leeS Y, chieh TS, hyr JM et al. prevalence and accessory gene regulator analysis of vancomycin intermediate staphylococcus aureus among MR isolates in Taiwan smart program, 2003. Eur J clinical microbial infect Dis 2010;29:383-9
- [7]. Richter SS, Satola SW, Crispall EK, Heilmann KP, Dohrn CL, Riahi F et al. Detection of staphylococcus aureus isolates with heterogeneous intermediate level resistance to vancomycin in US. J CLINICAL MICROBIAL 2011;49:4203-7
- [8]. Hu J, Maxx Tian Y, Pang L, Cui LZ, Shan H reduced vancomycin susceptibility found in methicillin resistance and methicillin sensitive staphylococcus aureus clinical isolates in northern China. PLoS one 2013;08:e73300.
- [9]. Song JH, Hiramatsu K, Suh JY, Ko KS, Ito T, Kapi M et al. emergence in Asian countries of staphylococcus aureus with reduced susceptibility to vancomycin. Antimicrobial agents chemother 2004;48:40266-8.
- [10]. Lina G, Piemont Y, Godail-Gamat F, Bes M, Peter MO, Guaduchan V et al. Involvement of Pantone-valentine Leukocidine-Producing staphylococcus aureus in primary skin infection and pneumonia. Clin Infect Dis 1999;29:1128-32.
- [11]. Chang S, Sievert DM, Hageman JC, Boulton ML, Tenover FC, Downes FR, et al. Infection with VRSA containing the VanA resistance gene N Engl J Med 2003;348:1342-7.
- [12]. Tenover FC, Weingel LM, Appelbaum PC, Mc Dougal LK, Chaitram J, Mc Allister S, et al. VRSA isolates from a patient in Pennsylvania, antimicrobial agents chemother 2004;48:275-80.
- [13]. Saha B, Singh AK, Ghosh A, Bal M identification and characterization of a VRSA isolated from Kolkata (South Asia) Med Microbiol 2008;57:72-9.
- [14]. Shindia AA, Ragab FA, Nasrat HM, emergence of high level VRSA in the critical care patient care university hospitals and J basic and applied science 2011;5:1281-90.
- [15]. Clinical and Laboratory Standard Institute (CLSI): Performance standards for antimicrobial susceptibility testing. 18th International Supplement. CLSI document M 100-18. Wayne, Pennsylvania; 2008.
- [16]. Shilpa Rao. Textbook of microbiology Anantnarayana 6th edition.
- [17]. Muralidharan S. Special article on methicillin resistant *Staphylococcus aureus*. J Acad Clin Microbiol. 2009;11:15-6.
- [18]. Anupurba S, Sen MR, Nath G, Sharma BM, Gulati AK, Mohapatra TM. Prevalence of methicillin resistant *Staphylococcus aureus* in a tertiary referral hospital in eastern Uttar Pradesh. Indian J Med Microbiol. 2003;21:49-51
- [19]. Tiwari HK, Sen MR. Emergence of vancomycin resistant *Staphylococcus aureus* (VRSA) from a tertiary care hospital from northern part of India. BMC Infect Dis. 2006;6:156.
- [20]. Rajadurai Pandi K, Mani KR, Panneerselvam K, Mani M, Bhaskar M, Manikandan M. Prevalence and antimicrobial susceptibility pattern of methicillin resistant *Staphylococcus aureus*: A multicentre study. Indian J Med Microbiol. 2006;24:34-8.
- [21]. Majumder D, Bordoloi JS, Phukan AC, Mahanta J. Antimicrobial susceptibility pattern among methicillin resistant *staphylococcus* isolates in Assam. Indian J Med Microbiol. 2001;19:138-40
- [22]. Haq JA, Rahman MM, Asna SM, Hassain MA, Ahmed I, Haq T et al. MRSA in Bangladesh a multicenter study. Int J Antimicrob agents 2005;25:276-7.
- [23]. Taj Y, Abdullah FE, Kazmi SU, current pattern of antibiotic resistance in SA clinical isolates and emergence of Vancomycin resistance J. physician surg Pak 2010;20:728-32.
- [24]. Hafeez R, Chuhtai AS, Aslam M, prevalence and antimicrobial susceptibility of MRSA Int J Pathol 2004;2:10-5.
- [25]. Kaur H, Purwar S, Saini A, Kaur, Karadesai SG, Khdkute SD et al. status of MRSA infection and evaluation of PVL producing strains in Belgaum, JKIMSU 2012;1:43-51.

MR. RAVI VASHISTHA. "Prevalence and Antimicrobial Susceptibility pattern of Methicillin Resistant Staphylococcus aureus in Different Clinical Infections." IOSR Journal of Dental and Medical Sciences (IOSR-JDMS), vol. 18, no. 2, 2019, pp 31-33.