

## MicroRNA in Ovarian Cancer: A short review

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**Abstract:** Ovarian cancer represents the most fatal among gynaecological malignancies. The high mortality rate may be due to its late-stage diagnosis in lack of relevant diagnostic markers for early detection. There is a strong need for biomarkers that facilitate detection at an early stage. MicroRNAs (miRNAs), representing a new class of biomarkers are being explored. They are single-stranded short sequence RNAs that do not encode proteins but regulate target genes post-transcriptionally. They play a role as suppressors and promoters of ovarian carcinoma being involved in growth, inhibition of apoptosis, metastasis, invasion, and angiogenesis. The research done in this field has shown that miRNAs can facilitate discrimination of patients with ovarian carcinoma from healthy controls suggesting their use as diagnostic biomarkers. This review will summarize the current knowledge and clinical relevance of circulating miRNAs supporting their use in early diagnosis and prognosis of ovarian cancer.

**Keywords:** Circulating miRNA, Ovarian cancer, diagnosis, prognosis

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### I. Introduction

Ovarian carcinoma is lethal among malignancies of female reproductive system with incidence rate of approximately 2,40,000 diagnosed per year.<sup>1</sup> Majority of Ovarian carcinoma (approximately 90%), are epithelial in origin with five most common morphological subtypes such as high grade and low grade serous constituting 70% and 5% respectively, mucinous (3%), endometrioid and clear-cell (10%) each, besides others. Serous carcinoma is the most common subtype constituting approximately 80% of all ovarian tumors.<sup>2,3</sup> There are no specific clinical symptoms in early stage so, often detected at a late stage presenting with metastasis and invasion and 5- year life expectancy rate of 42.9% approx. Over 80% of advanced ovarian cancer patients shows relapse, representing poor prognosis.<sup>4,5</sup> Estimation of serum CA-125 and Transvaginal ultrasound (TVUS) is being done to diagnose Ovarian cancer. But, they do not increase viability in symptomless women with no genetic risk mutation. Instead, ovarian cancer screening with CA-125 and TVUS often carries the risk of false positive results leading to surgeries and associated complications that may lead even to death.<sup>6</sup> CA-125 has limited diagnostic sensitivity and specificity.<sup>7</sup> Studies are undergoing to find out molecular alterations that are occurring in ovarian cancer. So, that better diagnostic strategies for early diagnosis can be find out. In this regard, microRNAs, the small non-coding RNA of 19-25 nucleotides representing next-generation biomarkers are being investigated for diagnosis of ovarian cancer. miRNAs are found to increase or decrease their expression in some cancer types. miRNAs are regulating expression of more than half of the protein coding genes in human constituting approximately 60%.<sup>8</sup> So, are impacting natural processes such as cellular growth, cell differentiation, metabolism, ageing, inflammation and immune response. In this way, miRNA are involved in development and advancement of the tumor.<sup>9-13</sup> The review will summarize association of miRNA with various aspects of ovarian cancer.

### BIOGENESIS OF MICRORNA

microRNA precursors are in clusters and are found in many portions of the human genome. They are most frequently found within stretches of DNA sequences between the genes and intervening sequences within coding sequence of protein coding genes. These regions are basically referred as junk DNA as their function is unknown. They are infrequently found in exons of RNA transcripts and antisense transcripts.<sup>14,15</sup>

miRNAs are transcribed by RNA II polymerase and Initially, long primary miRNAs (Pri-miRNA) are formed. Primary miRNA has 33bp stem and terminal loop with flanking segments.<sup>16,17</sup> Drosha, required for processing has two domains consisting of RNA polymerase III and ds RNA binding component.<sup>18-20</sup> Within nucleus, Drosha in association with a microprocessor complex including DGCR8, KSRP, P68 proteins

processes pri-miRNA to form 70 nucleotide precursor microRNA (pre-miRNA)<sup>16,17</sup> Drosha cleaves both the ends of pri-miRNA and form two nucleotide sticky ends at 3' end.<sup>18-20</sup> Exportin-5, a Ran GTP dependent dsRNA binding protein transports pre-miRNA to cytoplasm where Dicer, a RNA specific ribonuclease processes it to miRNA duplex.<sup>16,17</sup> Factor like TRBP (Transactivating response RNA binding protein) is also required for regulating Dicer processing.<sup>21</sup> The two strands of miRNA uncoil and fully mature miRNA clusters into RNA induced silencing component (RISC). The miRNA functions as guiding strand and base pairs with selected mRNA.<sup>16,17</sup> The miRNA circulating in blood are in bound form with AGO<sub>2</sub>, a protein which is required for functional activity of RISC complex. With this complex, miRNA enter recipient cells and modulate gene expression. miRNA regulate the expression of genes at post transcription level by binding to 3'-UTR of target mRNA.<sup>22</sup> Binding of miRNA with high complementarity, leads to cleavage and degradation of mRNA target and if binds with low complementarity, inhibition of translation is the outcome.<sup>23</sup> The degradation of mRNA starts by canonical deadenylation machinery known as P-bodies that causes shortening of Poly A tail<sup>24,25</sup>

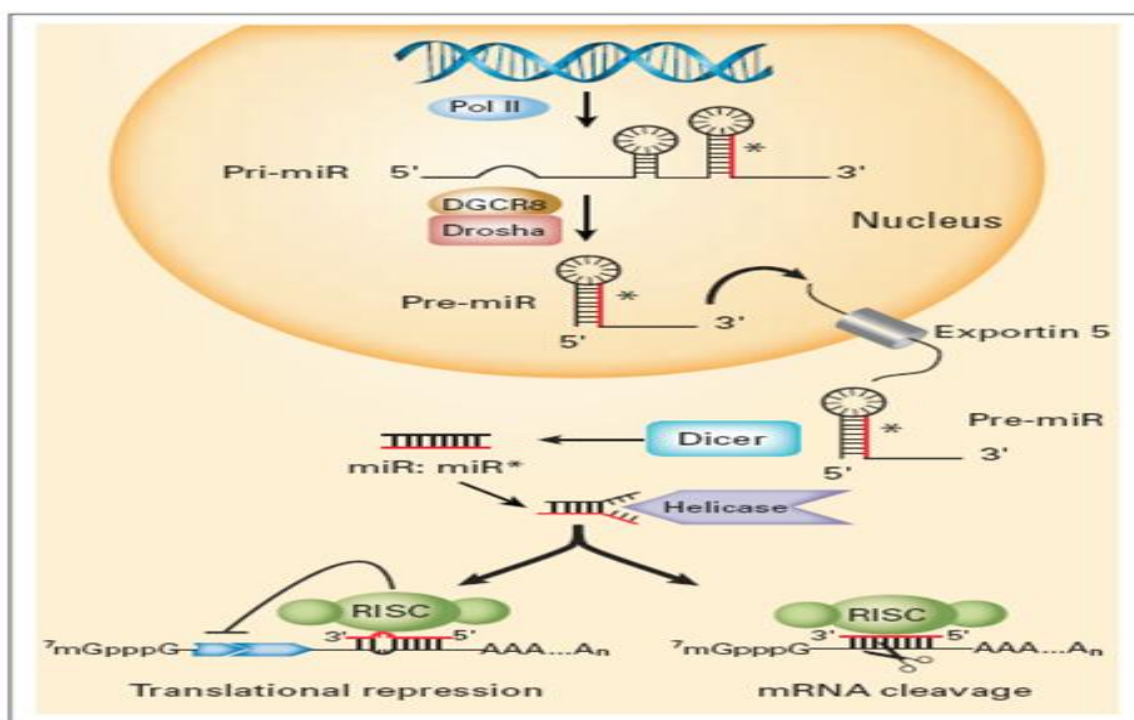


Figure 1: Biogenesis of microRNA<sup>26</sup>

### MOLECULAR DYSREGULATIONS OF MICRORNA IN OVARIAN CARCINOMA

Dysregulation of microRNAs is a common event in all stages and types of cancers. Explanation of this dysregulation is given by different mechanisms: alterations in chromosomes of miRNA gene like deletion, amplification, point mutations etc., variation in epigenetic mechanism regulating miRNAs, transcriptional modulation, variations in transcriptional and post transcriptional machinery in charge of microRNA formation<sup>26</sup>

The chromosomal alteration has been identified for several miRNA genes. About 50% of them are found at heritable point on a chromosome that may tend to break due to replication stress and at regions of amplification, deletion, or chromosome break points.<sup>27</sup> An elaboration of miR-182 region and removal of miR-15 region is found in ovarian carcinoma.<sup>28</sup> Loss of heterozygosity also contributes to ovarian cancer. The loss of normal copy of genes at loci let-7a-3/let-7b and miR-143/miR-145 has also been seen in 50 and 22% of 90 ovarian carcinoma respectively.<sup>29</sup> Zhang and colleagues established by genomic hybridization analysis of 227 human tumor samples that in, ovarian cancer, breast cancer and melanomas, certain specific fixed positions on a chromosome containing genes of miRNA were frequently altered. 26 miRNAs were found to be associated with increase in copies of the gene and 15 miRNAs with copy number loss of the gene.<sup>30</sup> High grade serous epithelial ovarian cancer bestows mutations in *Tp53*, *BRCA1* and *BRCA2*. Low grade serous epithelial ovarian cancer present with mutations in *KRAS*, *BRAF*, *PIK3CA*, *CTNNB1* and *PPP2RIA*.<sup>31,32</sup> TP53 mutations are seen in about 96% of high grade serous ovarian cancer. The expression of tumor suppressor miRNA 34a, 34b and 34c, was reduced 100% and 72% respectively with TP53 mutation.<sup>33</sup> The most of these genetic variations are missense, 30% are frameshift mutations, nonsense or splice junction variants leading to total inadequacy of p53

protein.<sup>34</sup> About one-fifth of subjects with ovarian carcinoma have hereditary disease related with germinal mutations of BRCA1 and BRCA2.<sup>35,36</sup> The variant allele of miR-146a is produced by G to C polymorphism of precursor miR-146a. It can increase its expression and modulates expression of BRCA1/2 by binding to 3'-UTR of BRCA1 mRNA.<sup>37</sup> Epigenetic alterations are an important reason for dysregulation in miRNA. Expression of certain miRNA genes is also regulated via epigenetic mechanism including CpG island methylation within promoter region.<sup>38</sup> Methylation is involved in regulating 11% of miRNA genes.<sup>39</sup> The decrease in expression of miR-34 in ovarian carcinoma had also been found due to hyper methylation of tumor suppressor genes 34a, 34b and 34c.<sup>40,41</sup> Iorio and colleagues suggested by his study on ovarian cell lines OVCAR3 that overexpressed miRNA like miR-21, miR-203 and miR-205 in ovarian cancer might be regulated by methylation.<sup>42</sup> Some miRNA are regulated by transcription control like miR-200 family. This is also operated by transcription control in ovarian carcinoma. ZEB1/2, transcription factors binds to promoters of miR-200 clusters and blocks transcription.<sup>43</sup> Expression of miR-17-92 cluster is operated by transcription factor c-Myc.<sup>44</sup> Drosha and Dicer mutation is another mechanism which explains dysregulation of miRNA in ovarian cancer. In 39% of ovarian cancer tissue analyzed, a 60% decrease of dicer mRNA and 51% decrease of drosha mRNA levels has been evaluated.<sup>45</sup> In high grade and high stage ovarian carcinoma, a marked down regulation of dicer expression has been seen.<sup>46</sup> A recurrent somatic missense mutation of DICER1 has been discovered in non-epithelial ovarian tumor. These mutations are highly prevalent (60%) in sertolileydig cell tumour.<sup>47</sup>

### **ROLE OF MICRORNA IN TUMORIGENESIS**

Many studies has been done so far to suggest the role of microRNA in ovarian carcinoma. microRNA may act as tumor suppressor genes or oncogenes or involved in biological behaviour in ovarian cancer development. They are also involved in invasion and metastasis of ovarian cancer cells by promoting epithelial to mesenchymal transition (EMT), angiogenesis, regulating extracellular matrix (ECM) or by regulating cellular growth factors. There is down regulation of expression of certain miRNA in tumor tissues suggesting their role as tumor suppressors. Tumor suppressors genes are known to inhibit tumor formation by inhibiting proliferation of cells, promoting differentiation and inhibiting migration of cells. miRNA may serve as proto-oncogenes which are involved in cell growth, reproduction, proliferation, differentiation. Certain miRNA shows significant up regulation in their expression in tumor tissues suggesting their role as proto-oncogenes. Their up regulation lead to unusual cell behavior and malignant growth of cells.

Luo et.al indicated tumor suppressor like effect of miR-126 on SKOV3 cell lines in ovarian cancer. It acts by inhibiting expression of PAK4.<sup>48</sup> miR-1 /miR-133a has a tumor suppressor like role in endometrial cancer. They regulate phosphodiesterase 7A (PDE7A) and bring inhibition of metastasis and invasion. Its levels are significantly downregulated in endometrial cancer.<sup>49</sup> Let -7 family hinders growth and invasion of tumor cells by inhibiting expression of proteins such as RAS, c-Myc, HMG-A2, c-Dc 25A, Cdk 6, cyclin-2 encoded by proto-oncogenes. The ovarian cells with more invasion and metastatic potential seems to have low let-7f expression.<sup>50</sup>

Fan et al. found that inhibition of miR-20a subdues the invasion by acting on Amyloid precursor protein (APP).<sup>51</sup> Zou et al. established that increased expression of miR-197 is related with tumor invasion and metastasis in OC by targeting nemo like kinase (NLK) and bringing its downregulation.<sup>52</sup> Depending upon cellular context miRNA may act as tumor suppressor or oncogene like miR-429 levels are increased initially in epithelial ovarian cancer but decrease with distant migration and metastasis.<sup>53</sup> Tumor cells metastasize by promoting EMT. EMT induces changes in cells like loss of morphological characteristics, reorganization of cytoskeleton and attainment of motile phenotype.<sup>54</sup> miR-200 family participate in epithelial to mesenchymal transition and invasion by targeting E-cadherin transcription repressors like ZEB1/ZEB2 to increase E-cadherin expression and in turn epithelial phenotype.<sup>55</sup> It also targets transcription factor snail. That also leads to enhanced E-cadherin expression in ovarian carcinoma.<sup>56</sup> E-cadherin is a glycoprotein that in normal circumstances helps in cell-cell adhesion along with  $\beta$ -catenin.<sup>57</sup> Tumor cells after getting induced by TGF- $\beta$  or PDGF-D increase ZEB1/ZEB2 expression and as a result decrease in miR-200 promoting EMT.<sup>58</sup> Dong et.al found that miR-137 and miR-34a function as unmediated suppressors of Snail (zinc finger transcription factor) in OC cells and suppress EMT phenotype and sphere formation of OC cells.<sup>59</sup> miR-205 shows increase in late stage of ovarian cancer.<sup>60</sup> It inhibits TCF-21 and increase ability of ovarian cancer cells to metastasize.<sup>61</sup> miR-205 down regulates Ezrin and lamin A/C after stimulation by VEGF resulting in proliferation, invasiveness and inhibition of apoptosis.<sup>62</sup> Overexpression of miR-125, miR-181b is related with invasion and metastasis of ovarian cancer cells by acting on large tumor repressor 2 (LATS2).<sup>63,64</sup> Li et.al established that in OC cells with low metastatic capability expression of miR-183 and miR-22 is more compared to ovarian cells with high metastatic potential. Rise in expression of miR-183 and miR-22 decreases the expression of Ezrin protein and in ezrin mediated way hinders ovarian cancer metastasis.<sup>65</sup> miR-543 causes transcription inhibition of MMP-7 mRNA by binding to its 3'-UTR and reduces metastasis and invasion.<sup>66</sup> MMPs are proteases involved in tumor invasion by cleaving components of ECM. Their increased expression is related with advancement

from benign to malignant ovarian cells.<sup>67</sup> They modulate cell adhesion molecules (CAM), growth factors(GF) and their receptors.<sup>68</sup> Angiogenesis is a physiological process required for cancer succession and metastasis by supplying oxygen and required nutrients. It is represented by continuous growth of new blood vessels from preexisting ones. Blood supply is crucial for cancer growth.<sup>69</sup> PTEN(Phosphatase and tensin homolog) is a tumor suppressor gene that mediates PI3K/AKT pathway required for normal blood vessel development.<sup>70</sup> PTEN converts phosphatidyl Inositol (3,4,5) triphosphate into Phosphatidyl inositol (4,5) bisphosphate.The PI3K deregulation causes activation of AKT.Activation of AKT signaling results in unrestricted proliferation and neoplastic angiogenesis.PTEN is required for stoppage of AKT signaling induced by oncogenes. miR-222 also targets PTEN and promote metastasis and invasion.<sup>71-73</sup> miR-205 induce angiogenesis by repressing PTEN and activating downstream AKT pathway.<sup>74</sup> Interleukin-8 (IL-8) and Chemokine Ligand-1(CXCL-1) secreted by tumor epithelial cells are main players of tumor vasculature and angiogenesis.miR-200 family targets IL-8 and CXCL-1 and inhibits formation of blood vessels.<sup>75</sup>Tumor angiogenesis also results from imbalance between proangiogenic and antiangiogenic factors specially of VEGF. In majority of ovarian carcinoma VEGF express at high levels.<sup>76</sup>Upregulated expression of miR-27a inhibits ZBTB 10 expression and in this way indirectly regulates expression of VEGF and VEGFR (receptor) leading to tumor growth and neoangiogenesis.<sup>77</sup>miR-125 b and miR-199 a acts as tumor suppressors and targets HIF-1 $\alpha$  and VEGF and decrease angiogenesis.<sup>78</sup> Xu et.al seen that miR-145 has a suppressing effect on neoangiogenesis and is found repressed in tissues and cell lineage of ovarian carcinoma.<sup>79</sup>Inhibition of apoptosis is also a reason of ovarian carcinoma.The apoptotic suppressor such as survivin member of Inhibitors of apoptosis (IAP) family has an inhibitory action on caspases cascade,The activators of apoptosis such as (Smac/Diablo) neutralizes the inhibitory action of IAPs on caspases and bring cellular apoptosis.Survivin suppress apoptosis by sequestration of Smac/Diablo.High serum survivin levels thus correlate with late stage, grade, ascites, peritoneal metastasis of serous ovarian cancer and level of cytoreduction.<sup>80-82</sup>

**MICRORNAs AS DIAGNOSTIC AND PROGNOSTIC BIOMARKERS**

In human,miRNAs present in circulation are stable and are protected from endogeneousRNase activity.<sup>83</sup>.Majority of miRNAs are released in body fluids via binding to lipoproteins like HDL or by forming non-vesicular Ago<sub>2</sub>ribonucleoprotein complexes and as small extracellular membranous vesicles called exosomes.<sup>22</sup> Their stability and specificity make them potential diagnostic biomarker in cancer. <sup>84</sup>In miRbase (version 21,june17), 2588 mature human miRNA and 1915 mature mouse miRNA have been identified.<sup>8</sup>miRNArepresents only ~ 0.01% of total RNA by weight. Average expression of individual miRNA species is around 500/cell that is greater than the expression of mRNA species.<sup>85</sup> Various studies has shown that microRNA can serve as diagnostic markers in ovarian cancer as they are related to various aspects of ovarian cancer occurrence.

Hausler et.al evaluated 24 patients of serous ovarian cancer and 15 normal healthy controls on microarray and found higher expression of miR 30c-1-3p and lower expression of miR181a-3p, miR450-5p, miR 342-3p in relapsed serous ovarian cancer group.<sup>86</sup>

Kan et.al performed expression profiling and found higher expression of miR-182,miR-200a,miR-200b,miR-200c in serous epithelial ovarian cancer cell lines in contrast to control group i.e normal human ovarian surface epithelial cells.miR-200a, miR-200b,miR-200c when individually normalized to serum volume and miR-103 showed greater expression in serum of serous epithelial ovarian cancer group. miR 200b and 200c when combined and normalized to miR-103 and serum volume were found to be positive classifiers of Serous epithelial ovarian cancer .<sup>87</sup>

Chung et.al suggested that miRNA can serve as a unique biomarker of epithelial ovarian cancer.He found underexpression of miR-132, miR-26a,miR-145 and let -7b in serous ovarian cancer group in contrast to controls.<sup>88</sup>Zheng et.al scanned miRNA by Taqman low density array and validated them by real time PCR assay.He found increased expression of miR-205 and under expression of let -7f in ovarian cancer group compared to controls .A combined use of miR-205 and let-7f can be helpful in diagnosis of EOC in patients of stage-1 disease.<sup>54</sup>

Zuberi et.al found higher expression of miR-200a, miR-200b,miR-200c in epithelial ovarian cancer group compared to controls.<sup>89</sup>

**Table 1: microRNAs Diagnostic biomarkers**

| Tumor type | Sample type | UpregulatedmiRNA           | Downregulated miRNA               | Control group    | References |
|------------|-------------|----------------------------|-----------------------------------|------------------|------------|
| SAC        | Whole blood | miR-30c-1-3p               | miR-181a-3p,miR-342-3p,miR-450-5p | Healthy controls | 86         |
| MAC, SAC   | Plasma      | miR-205                    | Let-7f                            | Healthy controls | 54         |
| MAC        | Serum       | miR-200a,miR-200b,miR-200c |                                   | Healthy controls | 89         |

|                                   |                |                                                 |                                  |                                       |    |
|-----------------------------------|----------------|-------------------------------------------------|----------------------------------|---------------------------------------|----|
| SAC                               | Serum          |                                                 | miR-132 miR-26a, let-7b, miR-145 | Healthy controls                      | 88 |
| SAC                               | Plasma         | miR-106b, miR-126, miR150,miR17,miR-20a,miR-92a |                                  | Benign neoplasms                      | 90 |
| SAC,MAC,CCAC,and EAC and other OC | Serum exosomes | miR-375,miR-1307                                |                                  | Healthy controls and benign neoplasms | 91 |

SAC:Serousadenocarcinoma,MAC:Mucinousadenocarcinoma,CCAC:Clear cell adenocarcinoma, EAC:Endometroidadenocarcinoma,OC:Ovarian carcinoma

Shapira et.al supported the fact that miRNA can differentiate women with Ovarian cancer from benign mass.They generated miRNA profiles from plasma of 42 women with SEOC,36 women with benign neoplasm and 23 age matched healthy controls.They found 22 miRNAs that showed differential expression between healthy controls and ovarian cancer group. Six miRNA profiles (miR-106b, miR-126, miR-150,miR-17,miR-20a,miR-92a)were able to differentiate between benign and OC group.<sup>90</sup>

Su et.al found up regulated expression of miR-375 and miR-1307 in serum exosomes of ovarian cancer patients in contrast to benign and healthy groups. An association of miR-1307 and miR-375 was found with tumor staging and lymph node metastasis respectively. Moreover, both have considerable prospective as targets of OC chemoresistance.<sup>91</sup>

Many studies have shown that miRNA can serve as biomarkers of prognosis in patients with ovarian cancer.

Zuberi et al.found association of up regulated expression of miR-200a with tumor histology and stage. Overexpression of miR-200c is related with metastasis to lymph nodes. miR-200a ,miR-200b and miR-200c were able to anticipate the prognosis and overall survival in epithelial ovarian cancer.<sup>89</sup>

**Table 2: MicroRNA as Prognostic biomarkers**

| Tumor type                                    | Sample type | Up regulated microRNA           | Down regulated microRNA | End point | References |
|-----------------------------------------------|-------------|---------------------------------|-------------------------|-----------|------------|
| SAC,MAC,CCAC,EAC and Other ovarian carcinomas | Serum       | miR-141                         | miR-200c                | OS        | 93         |
| SAC and others                                | Serum       | miR-221                         | -                       | OS        | 94         |
| MAC, SAC                                      | Plasma      |                                 | Let-7f                  | PFS       | 54         |
| MAC                                           | Serum       | miR-200a ,miR-200b and miR-200c |                         | OS        | 89         |

SAC: Serousadenocarcinoma, MAC:Mucinousadenocarcinoma,CCAC:Clear cell adenocarcinoma, EC:Endometroidadenocarcinoma,OC:ovarian carcinoma OS:Overall survival PFS:Progression free survival

Chen et.al performed Kaplan–meier overall survival analysis using TCGA data and showedmiRNA (hsa-miR135,miR-150,miR-340,miR-625,miR-1908,miR-31,miR-87,miR-96,miR-196b,miR-449 andmiR-1275) are related with high survival of patients with SOC.<sup>92</sup>

Gao et.al found that increased miR-200c expression is related with higher 2 year probability of survival and low miR-141 expression with significant higher survival rate.<sup>93</sup>

Zheng et.al indicated that decreased levels of let-7f might be predictive of unfavorable prognosis in EOC.<sup>54</sup> Hong et al. seen that overexpression of miR-221 come about as unfavorable self-sufficient factor of prognosis .It also has a role as diagnostic marker.<sup>94</sup>

## II. Conclusion

Ovarian cancer is relatively manageable if diagnosed at an early stage. The mechanism underlying its etiology is still incompletely understood. Till date, no reliable biomarker is known that can assist in early stage diagnosis. miRNAs are related to various aspects of ovarian cancer and are stable in circulation. They behave as oncomirs or tumor suppressors by regulating cell proliferation, metastasis, invasion, angiogenesis and apoptosis. The emerging evidences have suggested that circulating miRNAs hold great capability as upcoming non-invasive diagnostic and prognostic biomarkers. It has been seen in different studies that miRNAs even from the same tumor type shows inconsistent pattern. This might be due to lack of standardized protocols of selection of internal controls. Further studies at large scale with standardized protocols are required to include miRNAs in current testing regime for early detection and screening of ovarian cancer.

## References

- [1]. Webb PM, Jordan SJ. Epidemiology of epithelial ovarian cancer. *Best Pract Res ClinObstetGynaecol.* 2016. <https://doi.org/10.1016/j.bpobgyn.2016.08.006>
- [2]. Prat J. FIGO Committee on Gynecologic Oncology . Staging classification for cancer of the ovary, fallopian tube, and peritoneum. *Int J Gynaecol Obstet.* 2014;124:1–5.
- [3]. Prat J. New insights into ovarian cancer pathology. *Ann Oncol.* 2012;23 10:x111–x117.
- [4]. Reid BM., Permut JB., Sellers TA. Epidemiology of ovarian cancer: A review. *Cancer Biol. Med.* 2017; 14: 9–32.
- [5]. Matulonis UA, Sood AK, Fallowfield L, et al. Ovarian cancer. *Nat. Rev. Dis. Prim.* 2016;2:16061.
- [6]. Moyer VA. Screening for ovarian cancer: U.S. Preventive Services Task Force reaffirmation recommendation statement. *Ann Intern Med* 2012; 157: 900–904.
- [7]. Buys SS, Partridge E, Black A, Johnson CC, Lamerato L, Isaacs C. Effect of screening on ovarian cancer mortality: the Prostate, Lung, Colorectal and Ovarian (PLCO) cancer screening randomized controlled trial. *JAMA.* 2011;305:2295–302.
- [8]. Friedman RC, Farh KK, Burge CB, Bartel DP. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res.* 2009; 19:92–105. <https://doi.org/10.1101/gr.082701.108>.
- [9]. Hayes J, Peruzzi PP, Lawler S. MicroRNAs in cancer: Biomarkers, functions and therapy. *Trends Mol. Med.* 2014;20: 460–469.
- [10]. Sokol NS. Small temporal RNAs in animal development. *Curr. Opin. Genet. Dev.* 2012; 22: 368–373.
- [11]. He L, He X, Lim LP, de Stanchina E, Xuan Z et al. A microRNA component of the p53 tumour suppressor network. *Nature* 2007;447:1130–1134.
- [12]. Contreras J, Rao DS. MicroRNAs in inflammation and immune responses. *Leukemia* 2012; 26: 404–413.
- [13]. Wu Q, Yang Z, Shi Y, Fan D. MiRNAs in human cancers: The diagnostic and therapeutic implications. *Curr. Pharm. Des.* 2014; 20: 5336–5347.
- [14]. Lagos-Quintana M, Rauhut R, Meyer J, Borkhardt A, Tuschl T. New micro RNAs from mouse and human RNA. *2003;9:175-179.*
- [15]. Rodriguez A, Griffith-Jones S, Ashurst JL, Bradley A. Identification of mammalian microRNA host genes and transcription units. *Genome Res.* 2004;14:1902-1910.
- [16]. Krol J, Loedige I, Filipowicz, W. The widespread regulation of microRNA biogenesis, function and decay. *Nat. Rev. Genet.* 2010, 11, 597-610.
- [17]. Garofalo M, Croce CM. microRNAs: Master regulators as potential therapeutics in cancer. *Annu. Rev. Pharmacol Toxicol.* 2011; 51: 25-43
- [18]. Kim VN. MicroRNA biogenesis: Coordinated cropping and dicing. *Nat. Rev. Mol. Cell Biol.* 2005;6:376–385.
- [19]. Ha M, Kim VN. Regulation of microRNA biogenesis. *Nat. Rev. Mol. Cell Biol.* 2014;15:509–524.
- [20]. Han J, Lee Y, Yeom KH et al. Molecular basis for the recognition of primary microRNAs by the Drosha-DGCR8 complex. *Cell* 2006; 125: 887–901.
- [21]. Pratt AJ, Mac Rae II. The RNA-induced silencing complex: A versatile gene-silencing machine. *J. Biol. Chem.* 2009; 284: 17897–17901.
- [22]. Vasudevan S, Tong Y and Steitz JA: Switching from repression to activation: microRNAs can up-regulate translation. *Science.* 2007; 318: 1931-1934.
- [23]. Arroyo JD, Chevillet JR, Kroh EM, et al. Argonaute2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma. *Proc Natl Acad Sci U S A.* 2011;108:5003–8.
- [24]. Eulalio A, Behm-Ansmant I and Izaurralde E: P bodies: At the crossroads of post-transcriptional pathways. *Nat Rev Mol Cell Biol.* 2007 ;8: 9-22.
- [25]. SenG and Blau HM: Argonaute 2/RISC resides in sites of mammalian mRNA decay known as cytoplasmic bodies. *Nat Cell Biol.* 2005;7: 633-636.
- [26]. DiLeva G, Croce CM. Roles of small RNAs in tumor formation. *Trends Mol. Med.* 2010;16:257-267. [doi:10.1016/j.molmed.2010.04.001](https://doi.org/10.1016/j.molmed.2010.04.001).
- [27]. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noche E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci USA.* 2004;101:2999–3004.
- [28]. Zhang L, Volinia S, Bonome T, Calin GA, Greshock J, Yang N, et al. Genomic and epigenetic alterations deregulate microRNA expression in human epithelial ovarian cancer. *Proc Natl Acad Sci U S A.* 2008;105:7004–9.
- [29]. Bearfoot J, Choong DY, Goringe KL, Campbell IIG. Genetic analysis of cancer implicated microRNA in ovarian cancer. *Clin. Cancer Res.* 2008;14:7246-7250. [doi:10.1158/1078-0432.CCR-08-1348](https://doi.org/10.1158/1078-0432.CCR-08-1348).
- [30]. Zhang L, Huang J, Yang N, Greshock J, Megraw MS, Giannakakis A, et al. microRNAs exhibit high frequency genomic alterations in human cancer. *Proc Natl Acad Sci U S A* 2006;103:9136–41.
- [31]. Meinhold-Heerlein I, Bauerschlag D, Hilpert F, et al. Molecular and prognostic distinction between serous ovarian carcinomas of varying grade and malignant potential. *Oncogene.* 2005;24:1053–1065.
- [32]. Kurman R J, Shih J, et al. Molecular pathogenesis and extraovarian origin of epithelial ovarian cancer—shifting the paradigm. *Hum Pathol.* 2011;42:918–931.
- [33]. Salani R, Kurman RJ, Giuntoli R, Gardner G, Bristow R, Wang TL, et al. Assessment of TP 53 mutation using purified tissue samples of ovarian serous carcinomas reveals a higher mutation rate than previously reported and does not correlate with drug resistance. *Int J Gynecol cancer.* 2008;18:487-91.
- [34]. Lee CH, Subramanian S, Beck AH, et al. MicroRNA profiling of BRCA1/2 mutation-carrying and non-mutation carrying high-grade serous carcinomas of ovary. *PLoS ONE.* 2009; 4:e7314
- [35]. Miyamoto M, Takano M, Goto T, Kato M, et al. Clear cell histology as a poor prognostic factor for advanced epithelial ovarian cancer: a single institutional case series through central pathologic review. *J Gynecol Oncol.* 2013; 24: 37-43.
- [36]. Cannistra SA. Cancer of the ovary. *N Engl J Med.* 2004; 351: 2519-29.
- [37]. Pastrello C, Polesi J, Della Puppa L, Viel A, Maestro R. Association between hsa-mir-146a genotype and tumor age-of-onset in BRCA1/BRCA2-negative familial breast and ovarian cancer patients. *Carcinogenesis.* 2010;31: 2124–6.
- [38]. Lopez Serra P, and Esteller M. DNA methylation associated silencing of tumor suppressor miRNAs in cancer. *Oncogene.* 2012 ;31:1609-1622.
- [39]. Kunej T, Godnic I, Ferdin J, et al. Epigenetic regulation of miRNAs in cancer: an integrated review of literature. *Mutat. Res.* 2011;717: 7784.
- [40]. Vogt M, Munding J, Gruner M, et al. Frequent concomitant inactivation of miR-34a and miR-34b/c by CpG methylation in colorectal, pancreatic, mammary, ovarian, urothelial, and renal cell carcinomas and soft tissue sarcomas. *Virchows Arch* 2011;458:313–22
- [41]. Corney DC, Hwang CI, Matoso A, et al. Frequent downregulation of miR-34 family in human ovarian cancers. *Clin Cancer Res* 2010;16:1119–28.

- [42]. Iorio MV, Visone R, Di Leva G, et al. MicroRNA signatures in human ovarian cancer. *Cancer Res.* 2007;67:8699–707.
- [43]. Gregory PA, Bert AG, Peterson EL, et al. The miR-200 family and miR-205 regulates epithelial to mesenchymal transition by targeting ZEB1 and SIP1. *Nat. Cell Biol.* 2008;10:593-601. doi:10.1038/ncb1722
- [44]. Chang T C, Yu D, Lee YS., Wentzel EA., Arking DE, and West K M. Widespread miRNA repression by Myc contributes to tumorigenesis. *Nature Genet.* 2008; 40: 43-50.
- [45]. Meritt WM, Lin YG, Han IY, Kamat AA, et al. Dicer, Drosha and outcomes in patients with ovarian Cancer. *N Engl J Med.* 2008;359:2641-2650. doi:10.1056/NEJMoa0803785
- [46]. Pampalakis G, Diamandis EP, Katsaros D, Sotiropoulou G. Down regulation of dicer expression in ovarian cancer tissues. *Clin Biochem* 2010;43:324-327. doi:10.1016/j.clinchem.2009.09.014
- [47]. Heravi-Moussavi A, Anglesio MS, Cheng SW, et al. Recurrent somatic DICER1 mutations in nonepithelial ovarian cancer. *N Engl J Med.* 2012;366:234-242. doi:10.1056/NEJMoa1102903
- [48]. Luo P, Fei J, Zhou J, Zhang, W. microRNA-126 suppresses PAK4 expression in ovarian cancer SKOV3 cells. *Oncol. Lett.* 2015; 9:2225–2229.
- [49]. Yamamoto N, Nishikawa R, Chiyomaru T, et al. The tumor-suppressive microRNA-1/133a cluster targets PDE7A and inhibits cancer cell migration and invasion in endometrial cancer. *Int. J. Oncol.* 2015; 47: 325–334.
- [50]. Bussing I, Slack FJ, Grosshans H. let-7 microRNAs in development, stem cells and cancer. *Trends Mol. Med.* 2008; 14: 400–409.
- [51]. Fan X, Liu Y, Jiang J, et al. miR-20a promotes proliferation and invasion by targeting APP in human ovarian cancer cells. *Acta Biochim Biophys Sin (Shanghai)* 2010; 42: 318-324.
- [52]. Zou D, Wang D, Li R, et al. MiR-197 induces Taxol resistance in human ovarian cancer cells by regulating NIK. *Tumour Biol.* 2015; 36: 6725-6732.
- [53]. Meng X, Joosse SA, Müller V et al. Diagnostic and prognostic potential of serum miR-7, miR-16, miR-25, miR-93, miR-182, miR-376a and miR-429 in ovarian cancer patients. *Br J Cancer* 2015; 113: 1358-1366.
- [54]. Zheng H, Zhang L, Zhao Y, Yang D, Song F, Wen Y, et al. Plasma miRNAs as diagnostic and prognostic biomarkers for ovarian cancer. *PLoS One.* 2013;8:e77853.
- [55]. Park SM, Gaur AB, Lengyel E, Peter ME. The miR-200 family determines the epithelial phenotype of cancer cells by targeting the E-cadherin repressors zeb1 and zeb2. *Genes Dev.* 2008; 22:894–907.
- [56]. Lam SS, Mak AS, Yam JW, et al. Targeting estrogen-related receptor alpha inhibits epithelial-to-mesenchymal transition and stem cell properties of ovarian cancer cells. *Mol. Ther.* 2014; 22: 743–751.
- [57]. Huber MA, Kraut N and Beug H: Molecular requirements for epithelial-mesenchymal transition during tumor progression. *Curr Opin Cell Biol.* 2005;17: 548-558.
- [58]. Di Leva G and Croce CM. The role of microRNAs in the tumorigenesis of ovarian cancer. *Front. Oncol.* 2013;3:153. doi: 10.3389/fonc.2013.00153
- [59]. Dong et al. MiR-137 and miR-34a directly target Snail and inhibit EMT, invasion and sphere-forming ability of ovarian cancer cells. *Journal of Experimental & Clinical Cancer Research* 2016 ;35:132 DOI 10.1186/s13046-016-0415-y
- [60]. Greene SB, Gunaratne PH, Hammond SM, Rosen JM A putative role for microRNA-205 in mammary epithelial cell progenitors. *J Cell Sci* 2010;123: 606–618
- [61]. Wei J, Zhang L, Li J et al. microRNA-205 promotes cell invasion by repressing TCF21 in human ovarian cancer. *J. Ovarian Res.* 2017; 10: 33. [CrossRef]
- [62]. Li J, Li L, Li Z, Gong, G. et al. The role of miR-205 in the VEGF-mediated promotion of human ovarian cancer cell invasion. *Gynecol. Oncol.* 2015; 137: 125–133.
- [63]. Feng S, Pan W, Jin Y and zheng J. MiR-125 promotes ovarian cancer proliferation and motility by targeting IATS2. *Tumour Biol.* 2014; 35: 12339-12344.
- [64]. Xia Y and Gao Y: MicroRNA-181b promotes ovarian cancer cell growth and invasion by targeting IATS2. *Biochem Biophys Res Commun* 2014; 447: 446-451.
- [65]. Li J, Liang SH, Lu X. Potential role of ezrin and its related microRNA in ovarian cancer invasion and metastasis. *Zhonghua Fu chan Ke za Zhi.* 2010;45:787-792.
- [66]. Song N, Liu H, Ma X, Zhang S. Placental growth factor promotes metastasis of ovarian cancer through Mir-543 regulated MMP7. *Cell. Physiol. Biochem* 2015; 37:1104-1112.
- [67]. Schmalfeldt B, Prechtel D, Härtling K et al. Increased expression of matrix metalloproteinases (MMP)-2, MMP9 and the urokinase-type plasminogen activator is associated with progression from benign to advanced ovarian cancer. *Clin Cancer Res* 2001; 7:2396-2404.
- [68]. Overall CM and Kleinfeld O: Tumor microenvironment – opinion: validating matrix metalloproteinases as drug targets and antitargets for cancer therapy. *Nat Rev Cancer* 2006;6: 227-239.
- [69]. Folkman J. Tumour angiogenesis: therapeutic implications. *New Engl J Med.* 1971; 285: 1182-1186.
- [70]. Okumura N, Yoshida H, Kitagishi Y, Murakami M, Nishimura Y, Matsuda S. PI3K/AKT/PTEN Signaling as a Molecular Target in Leukemia Angiogenesis. *Adv Hematol.* 2012; 2012: 843085
- [71]. Bai Y, Yang Y, Yan Y, Zhong J, Blee AM, Pan Y, et al. RUNX2 overexpression and PTEN haplo insufficiency cooperate to promote CXCR7 expression and cellular trafficking, AKT hyperactivation and prostate tumorigenesis. *Theranostics.* 2019; 9: 3459-3475.
- [72]. Xue L, Huang J, Zhang T, Wang X, Fu J, Geng Z, et al. PTEN inhibition enhances angiogenesis in an in vitro model of ischemic injury by promoting Akt phosphorylation and subsequent hypoxia inducible factor-1alpha upregulation. *Metab Brain Dis.* 2018; 33: 1679-1688. 38
- [73]. Li B, Lu Y, Wang H, Han X, et al: miR-221/222 enhance the tumorigenicity of human breast cancer stem cells via modulation of PTEN/Akt pathway. *Biomed Pharmacother* 2016;79: 93-101.
- [74]. He L, Zhu W, Chen Q, et al. Ovarian cancer cell-secreted exosomal miR-205 promotes metastasis by inducing angiogenesis. *Theranostics* 2019; 9(26): 8206-8220. doi: 10.7150/thno.37455
- [75]. Merritt WM., Lin YG, Spann WA et al. Effect of interleukin-8 gene silencing with liposome-encapsulated small interfering RNA on ovarian cancer cell growth. *J. Natl. Cancer Inst.* 2008; 100: 359–372
- [76]. Yu L, Deng L, Li J, Zhang Y, Hu L. The prognostic value of vascular endothelial growth factor in ovarian cancer: a systematic review and meta-analysis. *Gynecol Oncol.* 2013;128(2):391–6.
- [77]. Lai Y, Zhang X, Zhang Z, Shu Y, Luo X, Yang Y, et al. The microRNA-27a-ZBTB10 specificity protein pathway is involved in follicle stimulating hormone-induced VEGF, Cox2 and survivin expression in ovarian epithelial cancer cells. *Int J Oncol* 2013;42(2):776–84. doi:10.3892/ijo.2012.1743 20.

- [78]. He J, Jing Y, Li W, Qian X, Xu Q, Li FS, et al. Roles and mechanism of miR199a and miR-125b in tumor angiogenesis. *PLoS One* 2013;8(2):e56647. doi:10.1371/journal.pone.0056647
- [79]. Q Xu, L.-Z. Liu X. Qian et al. MiR-145 directly targets p70S6K1 in cancer cells to inhibit tumor growth and angiogenesis. *Nucleic Acids Research* 2012; vol.40, no.2, pp.761–774.
- [80]. Martinez-Ruiz G, Maldonado V, Ceballos-Cancino G, Grajeda JP, Melendez-Zajgla J. Role of Smac/DIABLO in cancer progression. *J Exp Clin Cancer Res*. 2008;27:48. doi:10.1186/1756-9966-27-48.
- [81]. No JH, Jeon YT, Kim YB, Song YS. Quantitative detection of serum surviving and its relationship with prognostic factors in ovarian cancer. *Gynecol Obstet Invest*. 2011;71(2):136–40.
- [82]. Song Z, Yao X, Wu M. Direct interaction between surviving and Smac/ DIABLO is essential for the anti-apoptotic activity of surviving during taxol-induced apoptosis. *J Biol Chem*. 2003;278(25):23130–40.
- [83]. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A*. 2008;105:10513–8.
- [84]. Schwarzenbach H, Nishida N, Calin GA, Pantel K. Clinical relevance of circulating cell-free microRNAs in cancer. *Nat Rev Clin Oncol*. 2014;11:145–56.
- [85]. Regan C, Zuker M. *PLOS computational biology*. 2011;7:e1001090 (pubmed:21390282)
- [86]. Häusler SF, Keller A, Chandran PA, Ziegler K, Zipp K, Heuer S, et al. Whole blood-derived miRNA profiles as potential new tools for ovarian cancer screening. *Br J Cancer*. 2010;103:693–700.
- [87]. Kan CW, Hahn MA, Gard GB, Maidens J, Huh JY, Marsh DJ, et al. Elevated levels of circulating microRNA-200 family members correlate with serous epithelial ovarian cancer. *BMC Cancer*. 2012;12:627.
- [88]. Chung YW, Bae HS, Song JY, Lee JK, Lee NW, Kim T, et al. Detection of microRNA as novel biomarkers of epithelial ovarian cancer from the serum of ovarian cancer patients. *Int J Gynecol Cancer*. 2013;23:673–9.
- [89]. Zuberi M, Mir R, Das J, Ahmad I, Javid J, Yadav P, et al. Expression of serum miR-200a, miR-200b, and miR-200c as candidate biomarkers in epithelial ovarian cancer and their association with clinicopathological features. *Clin Transl Oncol*. 2015;17:779–87
- [90]. Shapira I, Oswald M, Lovecchio J, Khalili H, Menzin A, Whyte J, et al. Circulating biomarkers for detection of ovarian cancer and predicting cancer outcomes. *Br J Cancer*. 2014;110:976–83.
- [91]. Su et al. Upregulated expression of serum exosomal miR-375 and miR-1307 enhance the diagnostic power of CA125 for ovarian cancer. *Journal of Ovarian Research*. 2019; 12:6 <https://doi.org/10.1186/s13048-018-0477-x>
- [92]. chen et al. Identification of core aberrantly expressed microRNAs in serous ovarian carcinoma. *Oncotarget*, 2018; Vol. 9, (No. 29), pp: 20451-20466
- [93]. Gao YC, Wu J. MicroRNA-200c and microRNA-141 as potential diagnostic and prognostic biomarkers for ovarian cancer. *Tumour Biol*. 2015;36:4843–50.
- [94]. Hong F, Li Y, Xu Y, Zhu L. Prognostic significance of serum microRNA-221 expression in human epithelial ovarian cancer. *J Int Med Res*. 2013;41:64–71.
- [95]. [Figuresource.microRNABiogenesis.https://memorangapp.com/flascards/66902/4.17](https://memorangapp.com/flascards/66902/4.17)

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