Qualitative and Quantitative analysis of Phytotochemicals in leaf extracts of *Centella asiatica* L.

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Abstract: Centella asiatica is one of the chief medicinal herbs used for treating skin problems, wound, nervous disorders etc. and is found throughout tropical and sub tropical regions of India up to an altitude of 600m. Centella asiatica contains asiatic acid, asiaticoside and madecassoside as major phytochemical constituents that are responsible for pharmacological value apart from being rich in flavonoids and terpenoids.

The present study was carried out on six solvent extracts of Centella asiatica to investigate the presence of medicinally important phytochemicals in their leaves. All the six extracts revealed the presence of various phytochemicals such as tannins, phlobatannins, saponins, terpinoids, diterpinoids, emodins, flavonoids, cardiac glycosides, anthraquinones, carotenoids, reducing sugars, alkaloids, anthocyanin, coumarins, steroids, phytosterols, phenol, fatty acids, proteins and amino acids. The leaves of Centella asiatica contained a significant amount of alkaloid, flavonoids, phenolic, saponins and tannin content. The amount of flavonoids was maximum (45.75mg/gm) followed by phenols (25.85mg/gm), alkaloids (17.75mg/gm), saponins (16.75mg/gm) and tannins (14.45mg/gm). The concentration of total alkaloids was maximum in distilled water extract (35.85mg/gm), followed by methanol extract (17.65mg/gm), ethanol extract (15.75mg/gm), petroleum ether extract (14.75mg/gm), acetone extract (12.45mg/gm) and benzene extract (11.75mg/gm). The concentration of total flavonoids was maximum in ethanol and methanol extracts (42.45mg/gm and 42.65mg/gm respectively), followed by distilled water extract (27.87mg/gm), benzene extract (13.35mg/gm), petroleum ether extract (14.65mg/gm) and acetone extract (13.55mg/gm). The amount of total phenol was maximum in ethanol and methanol extracts (15.35 and 15.45mg/gm respectively), followed by distilled water extract (12.33mg/gm), benzene and petroleum ether extracts (9.85 and 11.67mg/gm respectively) and acetone extracts (10.65mg/gm). Saponin concentration was maximum in ethanol (16.75mg/gm), benzene extract (15.25mg/gm) and distilled water extract (15.35mg/gm). Acetone and petroleum ether extracts contained relatively least amount of saponins (10.55 and 11.45mg/gm respectively). The total tannin concentration was maximum in ethanol extract (12.25mg/gm), followed by petroleum ether, acetone and benzene extracts (11.55, 10.35 and 10.35 mg/gm) respectively. Methanol and distilled water extracts contained relatively low amount of total tannins, 9.25 mg/gm and 9.45 mg/gm respectively.

The data obtained in the present study is expected to serve as valuable tool for identification, authentication and detection of adulterants, standardization and quality control of the drugs. Hence it can be concluded that the results of the present study have given qualitative and quantitative information about the purity standards of the leaves of Centella asiatica.

Key words: Centella asiatica, Phytochemicals, Acetone, Petroleum ether, Ethanol, Methanol, Benzene, Distilled water

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I. Introduction

Centella asiatica L. (Syn Hydrocotyle asiatica Linn.) belonging to family Umbelliferae/Apiaceae of dicotyledonous angiosperm is a medicinal herb in India, China, Srilanka, Nepal and Madagascar. *Centella asiatica* is one of the chief herbs for treating skin problems, to heal wounds, for revitalizing the nerves and brain cells, and hence it is known as a "Brain food" in India. This herb is also known as Indian Pennywort, Gotu Kola, Asiatic pennywort, Spade leaf and Brahmi.

In Southeast Asia, it is traditionally used for the treatment of a wide variety of disorders such as skin diseases, rheumatism, inflammation, syphilis, mental illness, epilepsy, hysteria, dehydration, and diarrhea (Shanghai, 1977; Yu *et al.*, 2006) [1, 2]. In Indian systems this plant is used as medicine for enhancing memory

and for the treatment of skin diseases and nervine disorders (Jamil *et al.*, 2007) [3]. The plant medicinal properties have long been utilized by the people of Java and Indonesia. In China, it is indigenously called as Gotu kola, and it was one of the documented "miracle elixirs of life" (Diwan *et al.*, 1991) [4]. Herbal medicines can be used as adaptogens, these plant derived drugs either reduce stress reactions in the alarm phase and provide a certain degree of safety against long-term stress (Wagner *et al.*, 1994) [5]. *C. asiatica* is used to treat various ailments across India which includes body aches, headaches, insanity, asthma, leprosy, ulcers, eczemas, and wound healing (Mishra, 2003) [6]. *Centella asiatica* is an important medicinal herb used in the orient (Bown, 1995) [7], and is also popular in the West (Chevallier, 1996) [8]. It has been used as a medicine in the Ayurvedic tradition of India for thousands of years and listed in the historic '*Sushruta Samhita*', an ancient Indian medical text (Chopra *et al.*, 1986; Diwan *et al.*, 1991) [9, 4]. This medicinal plant has been listed as Threatened plant species by the International Union for Conservation of Nature and Natural Resources (IUCN) (Pandey *et al.*, 1993) [10], and also as an endangered species (Singh, 1989; Sharma and Kumar, 1998) [11, 12].

Botanical profile: *Centella asiatica* (L.) is a prostrate, faintly aromatic, stoloniferous, perennial, creeper herb, attaining a height up to 15cm. Stem is glabrous, striated, rooting at the nodes. This plant flourishes extensively in shady, marshy, damp and wet places such as paddy fields, river banks forming a dense green carpet and rather than clayey soil, the sandy loam (60% sand) is found to be the most fertile soil for its regeneration (Devkota Anjana and Jha, 2009) [13]. The leaves, 1-3 from each node of stems, long petioled, 2-6cm long and 1.5-5cm wide, orbicular-renniform, sheathing leaf base, crenate margins, glabrous on both sides. Flowers are in fascicled umbels, each umbel consisting of 3-4 white to purple or pink flowers, flowering occurs in the month of April-June. Fruits are borne throughout the growing season. It is about 6.5 cm long, oblong, globular in shape and has strongly thickened pericarp. Seeds have pendulous embryo which are laterally compressed.

Centella asiatica is found throughout tropical and sub tropical regions of India up to an altitude of 600m. The plant has been reported to occur also at high altitudes of 1550m in Sikkim and 1200m in Mount Abu (Rajasthan). The plant is indigenous to South-East Asia, India, Sri- Lanka, parts of China, the Western South Sea Islands, Madagascar, South Africa, South East USA, Mexico, Venezuela, Columbia and Eastern South America (Subban Ravi *et al.*, 2008) [14].

Phytochemicals of *Centella asiatica:* Centella asiatica contains asiatic acid, asiaticoside and madecassoside as major phytochemical constituents that are responsible for pharmacological value apart from being rich in flavonoids and terpenoids (Roy *et al.*, 2013) [15]. Centelloid was term given for different constituents of secondary metabolites produced by plant which mainly comprised of pentacyclic triterpenoid saponins (James and Dubery, 2009) [16]. Centellin, Asiatic and centellicin are also present in the aerial part of the plant (Siddiqui *et al.*, 2007) [17]. From plant extract madecassoside, asiaticoside, madecassic acid and asiatic acid have been isolated in the significant amount (Inamdar et al., 1996) [18]. A quantitative estimation of triterpene and a saponin, $2\alpha, 3\beta, 23$ -trihydroxyurs-20-en-28-oic acid and $2\alpha, 3\beta, 23$ - trihydroxyurs-20-en-28-oic acid and $2\alpha, 3\beta, 23$ - trihydroxyurs-20-en-28-oic acid from the aerial part of *C. asiatica* (Yu *et al.*, 2007) [20]. Chemical structure of some major phytochemicals of *Centella asiatica* is illustrated below:



R1= OH, R2= H: Asiatic acid R1=OH, R2= O : Madecassic acid R1= O-glu-gluc-gluc, R2= H: Asiaticoside R1= O-glu-gluc-gluc, R2=OH: Madecassoside

Asiatic acid derivates



Pharmacological importance: The whole plant is used for medicinal purposes (Singh and Singh, 2002) [21]. It is widely used as a blood purifier as well as for treating high blood pressure, for memory enhancement and promoting longevity. In Ayurveda, *Centella asiatica* is one of the main herbs for revitalizing the nerves and brain cells. Eastern healers relied on this plant to treat emotional disorders, such as depression, that were thought to be rooted in physical problems (PDR for herbal medicine, 1999; Hagemann et al., 1996) [22, 23]. In the Western medicine, during the middle of the twentieth century, *Centella asiatica* and its alcohol extracts were used in the treatment of leprosy (Baily, 1945) [24]. *Centella asiatica* shows a wide range of pharmacological activities viz.

ANTICANCER ACTIVITY: Asiatic acid is the major phytochemical of Centella asiatica which shows anticancer activity, particularly against cell lines of human breast cancer (Wang *et al.*, 2013; Pittella *et al.*, 2009; Babykutty *et al.*, 2009; Hussin *et al.*, 2014; Wu *et al.*, 2017; Park *et al.*, 2005; Zhang *et al.*, 2013; Kwon *et al.*, 2014) [25-32].

ANTIBACTERIAL ACTIVITY: Methanol hot extract from *C. asiatica* leaves shows antibacterial activity against *Staphylococcus aureus* ATCC 25923, Bacillus subtilis, Escherichia coli, Bacillus cereus, Pseudomonas aeruginosa (Zaidan *et al.*, 2005; Oyedeji *et al.*, 2005; Pitinidhi Pat, 2015; Sekar *et al.*, 2011) [33-36].

ANTIFUNGAL ACTIVITY: The petroleum ether, ethanol, chloroform, n-hexane, and aqueous extract of *C. asiatica* shows antifungal activity against *Aspergillus niger, Candida. Albicans and Aspergillus flavus* (Dash *et al.*, 2011; Dhiman *et al.*, 2016; Idris *et al.*, 2017; Sultan *et al.*, 2014) [37-40].

ANTI-INFLAMMATORY ACTIVITY: Plants with medicinal properties are rich in ceramide and different forms of terpenoids which show anti-inflammatory activity (Prakash, 2017) [41]. Pentacyclic triterpenoid and saponins are collectively known as centelloids that are responsible for therapeutic actions. The Centella asiatica extract showed moderate anti-inflammatory property on prostaglandin E2-induced inflammation in a dose-dependent manner (Somchit *et al.*, 2004) [42]. The aqueous and alcoholic extract of *C. asiatica* showed inhibition of edema (George *et al.*, 2009) [43]. The asiatic acid reduced paw edema by regulation of catalase,

superoxide dismutase (SOD), and glutathione in the liver tissue (Huang *et al.*, 2011) [44]. It was observed that the methanolic extract showed significant inhibition of oedema (Saha *et al.*, 2013) [45].

NEUROPROTECTIVE ACTIVITY: Neuroprotection aspect of *C. asiatica* mainly involves enzyme inhibition, prevention of amyloid plaque formation in Alzheimer's disease, dopamine neurotoxicity in Parkinson's disease, and reducing oxidative stress (Orhan, 2012) [46]. Aqueous extract of *C. asiatica* was evaluated on the activity of subtypes of phospholipase A2 (PLA2) in primary cultures of rat cortical neurons, asiaticoside present in extract inhibited cPLA2 and sPLA2 activities (Defillipo *et al.*, 2012) [47]. In male Sprague-Dawley rats, improved learning and memory were observed on acute administration of asiatic acid (Nasir *et al.*, 2011) [48]. Neuroprotective potential of modern medicine constituents of the plant includes asiatic acid, madecassic acid, and brahmaside as well as flavonoids madecassoside and madesiatic acid (Thomas *et al.*, 2015) [49]. *C. asiatica* was explored for neuroprotective effect on cell death and cognitive irregulation in aluminum-treated rat. Significant improvement in memory performance, oxidative defense was observed on chronic administration of CA (Prakash and Kumar, 2013) [50]. The plant is known to utilize neuroprotective effects by attenuating the changes in an animal model such as pathological neurobehavioral and neurochemical properties. Phosphoinositides-assisted cytodynamics and synaptic function show the neuroprotective effects of asiaticoside in the rat which includes mode of ROT-infused hemiparkinsonism (Gopi *et al.*, 2017) [51].

ANTIOXIDANT PROPERTY: *C. asiatica* extract and powder was evaluated for reduction in oxidative stress in Sprague-Dawley rats. Results showed a decrease in the generation of ROS and oxidative stress in the rats. It was also noted that there was a significant decrease in SOD level (Hussin *et al.*, 2007) [52]. Essential oil of *C. asiatica* extracted through steam distillation showed to be excellent antioxidant for food containing lipids. Its activity was quite comparable with the synthetic antioxidant butylhydroxyanisole (BHA) (Raza *et al.*, 2009) [53]. Polyphenol, flavonoid, β -carotene, tannin, Vitamin C, and DPPH compounds are readily found in *C. asiatica* contributing to significantly higher antioxidant activity in the herb (Chandrika *et al.*, 2015) [54]. Crude methanolic extract on continuous supplementation for 14 days resulted in increase in level of antioxidant enzymes and ascorbic acid level reduced in lymphoma-bearing mice (Jayashree *et al.*, 2003) [55]. Extracts of *C. asiatica* in different solvents such as chloroform, hexane, acetone, ethyl acetate, methanol, and water were assessed for antioxidant potential. The DPPH and hydroxyl radical scavenging activity were tested for methanolic extract which showed the IC50 value of 0.07 mg/ml and 500 µg/ml, respectively (Anand *et al.*, 2010) [56].

WOUND HEALING: The extract of Centella asiatica showed wound healing activity in a number of experimental animals (Shetty *et al.*, 2006; Somboonwong *et al.*, 2012; Yao *et al.*, 2017) [57-59].

ANTIDEPRESSANT: Compared to diazepam *C. asiatica* possesses antianxiety effect but has no effect on behavioral despair. Total triterpenes and imipramine from *C. asiatica* were evaluated for antidepressant activity using forced swimming test, the result showed a reduction in stillness duration and regulated amino acid levels (Chen *et al.*, 2003; Kalshetty *et al.*, 2012; Ceremuga *et al.*, 2015) [60-62].

ANTIDIABETIC ACTIVITY: Antidiabetic properties of leaf extract of *C. asiatica* was evaluated in alloxaninduced rat model and showed reduction in blood glucose level (Rahman *et al.*, 2012) [63]. Effect of ethanol extract was tested in streptozotocin (50 mg/kg)-induced Wistar rats. Studying the serum glucose, urea cholesterol, lipid, liver glycogen level, and body weight, the antidiabetic activity of extract at concentration of 200 mg/kg was noticed (Gayathri *et al.*, 2011; Supkamonseni *et al.*, 2014; Haque *et al.*, 2013; Kabir *et al.*, 2014; Maulidiani *et al.*, 2016) [64-68]. Asiatic acid was found to reduce blood glucose level in Goto-Kakizaki (GK) rat by enhancing fibrosis of islets in diabetes which plays a vital role in the prevention of islets dysfunction (Wang *et al.*, 2015) [69]. In diabetic Wistar rat model, asiatic acid showed to preserve and restore beta cell mass (Liu *et al.*, 2010) [70].

COGNITIVE FUNCTION: Asiatic acid was found to prevent spatial working memory and reduction of neurogenesis defects in the hippocampal region caused by 5-FU chemotherapy (Chaisawang *et al.*, 2017) [71]. Water extract of *C. asiatica* was observed to enhance synaptic differentiation and dendritic arborization with reference to $A\beta$ which causes cognitive improvement (Gray *et al.*, 2017) [72]. In a study, gotu-kola extract was supplemented for weeks in defined concentration results showed to be effective in the treatment of cognitive function impairment after stroke (Farhana *et al.*, 2016) [73]. Asiatic acid has potential to restore the impairment of cell proliferation, spatial working memory caused by treatment with valproic acid (Umka *et al.*, 2016) [74]. Water extract helped to improve cognitive function by activation of antioxidant response gene and mitochondrial biogenesis (Gray *et al.*, 2016) [75], normalized calcium homeostasis (Gray *et al.*, 2015) [76].

HEPATOPROTECTIVE: Effect of methanolic extract of *Centella asiatica* was evaluated in Type 2 diabetes mellitus, and showed reduction in hepatic concentrations of interleukin-1 β , MCP-1, and tumor necrosis factor alpha in diabetic control rats (Oyenihi *et al.*, 2017) [77]. In dimethylnitrosamine-induced liver injury *C. asiatica* noticeably enhanced fibrosis of liver tissues by mass periportal±bridging necrosis, intralobular degeneration, and focal necrosis (Choi *et al.*, 2016; Ghosh *et al.*, 2017; Duggina *et al.*, 2015) [78-80]. Asiatic acid protects liver injury by onset of Smad7-dependent inhibition of TGF-beta/Smad-assisted fibrogenesis (Tang *et al.*, 2012) [81]. Conventionally, used plants to get rid of liver dysfunction might, therefore, could be potential source for new hepatoprotective compounds for development as pharmaceutical entities (Rajalingam *et al.*, 2016) [82].

The pharmacological activities of *Centella asiatica* can be summarized by a Diagram:



Diagram: Pharmacological activities of *Centella asiatica*

In the present investigation phytochemicals of leaves of *Centella asiatica* were analyzed qualitatively and quantitatively in six solvent extracts viz. Acetone, Petroleum ether, Ethanol, Benzene and Methanol and the Distilled water.

II. Materials and Methods

In the present investigation the dried leaves of *Centella asiatica* were powdered using a mixture grinder and stored in air-tight container for future use. Six different solvents (five non polar viz. Acetone, Petroleum ether, Ethanol, Benzene and Methanol and one polar solvent, the Distilled water) were used for preparation of solvent extracts. The dried plant sample was soaked separately with acetone, petroleum ether, ethanol, benzene, methanol and distilled water under reflux condition for the solvent extract preparation. About 1 gm of the dried sample of leaves was added respectively into the test tubes containing 5 ml of solvents, and was extracted at room temperature. In the present investigation the important phytochemicals of leaves of *Centella asiatica* have been qualitatively and quantitatively analyzed for alkaloids, flavonoids, tannins, saponins and total phenols.

Phytochemical Analysis: Phytochemicals in leaves of *Centella asiatica* were analyzed qualitatively and quantitatively in all the six solvent extracts

Qualitative Phytochemical Analysis

The extracts in all the six solvents of leaves of *Centella asiatica* were tested for the presence of biological compounds by using following standard methods.

Test for Carbohydrates

Fehling's test: Equal volume of Fehling A and Fehling B reagents were mixed together and 2ml of it was added to crude extract and gently boiled. A brick red precipitate appeared at the bottom of the test tube indicated the presence of reducing sugars.

Benedict's test: Crude extract when mixed with 2ml of Benedict's reagent and boiled, a reddish brown precipitate formed which indicated the presence of the carbohydrates.

Iodine test: Crude extract was mixed with 2ml of iodine solution. A dark blue or purple coloration indicated the presence of the carbohydrate.

Test for Phenols and Tannins

Crude extracts were mixed with 2ml of 2% solution of FeCl₃. A blue–green or black coloration indicated the presence of phenols and tannins.

Test for Flavonoid

Alkaline reagent test: Crude extracts were mixed with 2ml of 2% solution of NaOH. An intense yellow color was formed which turned colorless on addition of few drops of diluted acid which indicated the presence of flavonoids.

Test for Saponins (Frothing test): Crude extracts were mixed with 5ml of distilled water in a test tube and it was shaken vigorously. The formation of stable foam was taken as an indication for the presence of saponin.

Test for Glycosides

Liebermann's test: Crude extracts were mixed with each of 2ml of chloroform and 2ml of acetic acid. The mixture was cooled in ice. Carefully concentrated H_2SO_4 was added. A colour change from violet to blue to green indicated the presence of steroidal nucleus, i.e., glycone portion of glycoside.

Salkowski's test: Crude extracts were mixed with 2ml of chloroform. Then 2ml of concentrated H_2SO_4 was added carefully and shaken gently. A reddish brown color indicated the presence of steroidal ring, i.e., glycone portion of the glycoside.

Keller-kilani test (Cardiac Glycosides): Crude extracts were mixed with 2ml of glacial acetic acid containing 1-2 drops of 2% solution of FeCl₃. The mixture was then poured into another test tube containing 2ml of concentrated H_2SO_4 . A brown ring at the inter phase indicated the presence of cardiac glycoside.

Test for Alkaloids: The crude extract of all the six solvents was boiled in 10 ml methanol and filtered separately. 1% HCl was added followed by 6 drops of Dragendroff reagent, and the brownish-red precipitate was taken as evidence for the presence of alkaloids.

Phlobatannins: The deposition of a red precipitate denoted the presence of phlobatannins when crude extract of all the six solvent of plant material was dissolved in 10 ml of aqueous extract and few drops of 1% HCl were added in the boiling tube.

Anthraquinones: All the six solvent extracts of leaves were boiled in 10% HCl for 5 mins separately and the filtrate was allowed to cool. An equal volume of $CHCl_3$ with few drops of 10% NH_3 was added to the 2ml filtrate. The formation of rose-pink colour implies the presence of anthraquinones.

Quantitative estimation of phytochemicals

Determination of Alkaloids: Alkaloids content was measured by method suggested by Harborne (Harborne, 1973) [83]. A suspension was prepared by dispersing 5 gm of the dried leaves in 10% acetic acid solution in ethanol and kept at 28° C for 4hrs which was further filtered through Whatman No. 42. Thereafter alkaloid was precipitated by concentrating the filtrate to one-quarter of its original volume and drops of conc. aqueous NH₄OH were added. Finally, the precipitate was washed with 1% ammonia solution and dried at 80° C in the oven. The content of alkaloid was calculated and expressed as mg/g of sample.

Determination of Flavonoids: The flavonoids content was also determined by Harborne (Harborne, 1973) [83] method. 5 gm of leaves were boiled in 2M HCl for 30 min under reflux condition and filtered after cooling. An equal volume of ethyl acetate was then added drop wise to the filtrate. The weight of precipitated flavonoid was determined and recorded as mg/g.

Determination of Tannins: The finely powdered leaves of *Centella asiatica* were kept in a beaker containing 20 ml of 50% methanol covered with parafilm and then heated at 80oC in a water bath for 1 hr with continuous stirring. The extract was quantitatively filtered using a double layered Whatman No.1 filter paper and rinsed with 50% methanol. 1 ml of sample extract was treated with 20 ml distilled water, 2.5 ml Folin-Denis reagent and 10 ml of 17% Na₂CO₃ for the development of a bluish-green colour and was allowed to stand for 20 mins. The absorbance was measured at 760 nm and the amount of tannin was calculated by comparing it with a standard curve prepared in the range of 0-10 ppm.

Determination of Saponins: 100 ml Isobutyl alcohol was added to 1 gm of the finely powdered sample and stirred for 5 hrs. 20 ml of 40% saturated solution of Magnesium carbonate was added to the mixture and filtered. 2 ml of 5% FeCl3 solution and 50ml volume of distilled water was added to 1ml of colourless solution and kept for 30 mins for colour (blood red) development The absorbance of the samples as along with the standard were read at 380 nm and calculated in mg/g. Standard saponin solution was prepared in the reference range of 0-10 ppm.

Determination of total phenols: Five gms of the powdered leaves were boiled with 50 ml of ether for 15 mins and distributed in the ratio 1:2 (extract: distilled water). 2ml of ammonium hydroxide followed with 5ml of pentanol was added to it and incubated at the room temperature for 30mins. The absorbance was read at 505 nm wavelength.

For measuring alkaloids a suspension was prepared by dispersing 5 gm of the dried leaves in 10% acetic acid solution in ethanol and kept at 28° C for 4hrs which was further filtered through Whatman No. 42. Thereafter alkaloid was precipitated by concentrating the filtrate to one-quarter of its original volume and drops of conc. aqueous NH₄OH were added. Finally, the precipitate was washed with 1% ammonia solution and dried at 80° C in the oven. The content of alkaloid was calculated and expressed as mg/g of sample.

For determining flavonoids 5 gm of leaves were boiled in 2M HCl for 30 min under reflux and filtered after cooling. An equal volume of ethyl acetate was then added drop wise to the filtrate. The weight of precipitated flavonoid was determined and reported as mg/g.

For measuring tannin the finely powdered leaves of *Centella asiatica* were kept in a beaker containing 20 ml of 50% methanol covered with parafilm and then heated at 80° C in a water bath for 1 hr with continuous stirring. The extract was quantitatively filtered using a double layered Whatman No.1 filter paper and rinsed with 50% methanol. 1 ml of sample extract was treated with 20 ml distilled water, 2.5 ml Folin-Denis reagent and 10 ml of 17% Na₂CO₃ for the development of a bluish-green colour and was allowed to stand for 20 mins. The absorbance was measured at 760 nm and the amount of tannin was calculated by comparing it with a standard curve prepared in the range of 0-10 ppm.

For determining saponin content 100 ml Isobutyl alcohol was added to 1 gm of the finely powdered sample and stirred for 5 hrs. 20 ml of 40% saturated solution of Magnesium carbonate was added to the mixture and filtered. 2 ml of 5% FeCl₃ solution and 50ml volume of distilled water was added to 1ml of colourless solution and kept for 30 mins for colour (blood red) development The absorbance of the samples as along with the standard were read at 380 nm and calculated in mg/g. Standard saponin solution was prepared in the reference range of 0-10 ppm.

For determining total phenolic content five gms of the powdered leaves were boiled with 50 ml of ether for 15 mins and distributed in the ratio 1:2 (extract: distilled water). 2ml of ammonium hydroxide followed with 5ml of pentanol was added to it and incubated at the room temperature for 30mins. The absorbance was read at 505 nm wavelength.

Quantitative analysis of phytochemical constituents in six different solvent extracts

Six solvent extract of leaves of *Centella asiatica* viz. acetone, petroleum ether, ethanol, methanol, benzene and distilled water were prepared by soaking 10gm of the powdered sample in 200 ml of each of the solvent separately for 12 hrs. The extracts were then filtered using filter paper. The extracts were then concentrated to ¹/₄ of the original extracts i.e. 50 ml.

The amount of total phenolics in extracts was determined by the Folin–Ciocalteu method. Gallic acid was used as a standard by using different concentrations of (20-200µg) from which the total phenol content in the extract was expressed in terms of gallic acid equivalent (mg GAE /gm) extract. Different aliquots of 0.1 to 1.0 ml of plant extract were also prepared in methanol and 0.5 ml of each sample were introduced into test tubes and mixed with 2.5 ml of a 10-fold dilute Folin- Ciocalteu reagent and 2 ml of 7.5% sodium carbonate. The mixture was allowed to stand for 30 mins at room temperature. Phenols react with the phosphomolybdic acid in Folin-Ciocalteau reagent in alkaline medium and produce blue coloured complex (Molybdenum blue). The absorbance of the resulting solutions was measured at 760 nm against reagent blank. A standard calibration

curve was prepared by plotting absorbance against concentration and it was found to be linear over this concentration range. The concentration of total phenol in the test sample was determined from the calibration graph. The assay was carried out in triplicate and the mean values with \pm SD are presented.

The aluminium chloride colorimetric method was used for flavonoids determination. Each solvent extract (0.5 ml of 1:10 gm ml⁻¹) was separately mixed with 1.5 ml of methanol, 0.1 ml of 10% aluminium chloride, 0.1 ml of 1 M potassium acetate and 2.8 ml of distilled water. It was kept at room temperature for 30 min; the absorbance of the reaction mixture was measured at 418 nm. The percentage of total flavonoids were calculated from the calibration curve of Quercetin (200-1000 μ g) plotted by using the same procedure and total flavonoids was expressed as Quircetin equivalents (QE) in mg per gm sample.

The results obtained have been presented in Table-1, 2 and 3; Figure-1 and 2.

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Solvent	Т	Ph	Sa	Ter	Dte	Em	Fl	Ca	Ant	Cr	Re	Al	An	Со	St	Ps	Ph	F	Pr	Α
extracts	а	1	р	р	r	d	a	r	h	t	s	k	с	u	r	t	e	a	t	a
of																				
leaves																				
Acetone	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Petroleu	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+
m ether																				
Ethanol	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	-	+	+	+	+
Methan	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	-	+	+	+	+
ol																				
Benzene	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Distilled	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	-	+	+	+	+
water																				

 Table- 1: Phytochemicals of Centella asiatica analysed qualitatively in leaves in six different solvent extracts

Ta= Tannin; Phl= Phlobatannin; Sap= Saponin; Ter= Terpinoid; Dtr = Diterpinoid ; Emd= Emodin ; Fla= Flavonoid; Car= Cardiac glycoside; Anth= Anthraquinones; Crt= Carotenoids; Res= Reducing sugar; Alk= Alkaloid; Anc= Anthocyanin; Cou= Coumarin; Str= Steroids; Pstr= Phytosterol; Phe= Phenol; FA= Fatty acids; Prt= Protein; Aa= Aminoacids

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				-					

Phytochemicals	Amount in mg/gm
Alkaloids	17.75±0.25
Flavonoids	45.75±0.45
Phenols	25.85±0.65
Saponins	16.75±0.25
Tannins	14.45±0.25

Mean ± SD of five measurements



Figure-1: Phytochemicals observed in leaves of Centella asiatica

different solvent extracts of leaves of <i>Centella asiatica</i> (amount in mg/gm)										
Solvent Extracts	Total Alkaloids	Total Flavonoids	Total Phenol	Total Saponins	Total Tannin					
Acetone	12.45±0.35	13.55±0.43	10.65±0.65	10.55±0.34	10.35±0.17					
Petroleum ether	14.75±0.25	14.65±0.17	11.67±0.61	11.45±0.23	11.55±0.15					
Ethanol	15.75±0.35	42.45±0.15	15.35±0.24	16.75±0.21	12.25±0.21					
Methanol	17.65±0.17	42.65±0.16	15.45±0.21	13.38±0.31	9.25±0.12					
Benzene	11.75±0.16	13.35±0.41	9.85±0.19	15.25±0.41	10.35±0.16					
Distilled water	35.85±0.17	27.87±0.25	12.35±0.42	15.35±0.40	9.45±0.18					

Table- 3: Comparative analysis of total Alkaloids, Flavonoids, Phenol, Saponins and Tannins in six different solvent extracts of leaves of *Centella asiatica* (amount in mg/gm)

Tatal phenol in mg/gm is measured as Gallic Acid Equivalent (GAE/g extract); Total flavonoids in mg/gm is measured as Quercetin Equivalent (QE)/g extract.

Mean ± SD of five measurements



Figure-2: Comparative analysis of phytochemicals in six different solvent extracts of leaves of *Centella* asiatica

III. Results

The present study was carried out on six solvent extracts of *Centella asiatica* to investigate the presence of medicinally important phytochemicals in their leaves. All the six extracts revealed the presence of various phytochemicals such as tannins, phlobatannins, saponins, terpinoids, diterpinoids, emodins, flavonoids, cardiac glycosides, anthraquinones, carotenoids, reducing sugars, alkaloids, anthocyanin, coumarins, steroids, phytosterols, phenol, fatty acids, proteins and amino acids. Of these 20 phytochemicals tannin, saponin, terpenoid, diterpinoid, emodin, flavonoid, cardiac glycoside, anthraquinone, carotenoid, reducing sugar, alkaloids, anthocyanin, coumarin, phenol, fatty acid, protein and amino acids were detected in all the six solvent extracts. Phlobatannin, steroid and phytosterol were not detected in ethanol, methanol and distilled water extracts. Emodin was detected in all extracts except petroleum ether and benzene (Table-1).

From the results (Table- 2; Fig- 1) it is evident that the leaves of *Centella asiatica* contained a significant amount of alkaloid, flavonoids, phenolic, saponins and tannin content. The amount of flavonoids was maximum (45.75mg/gm) followed by phenols (25.85mg/gm), alkaloids (17.75mg/gm), saponins (16.75mg/gm) and tannins (14.45mg/gm) (Table- 2; Fig- 1).

The comparative analysis of phytochemicals viz. total alkaloids, flavonoids, phenols, saponins and tannins in six different solvent extracts from leaves of *Centella asiatica* has been presented in Table- 3 and Fig-2. From the results it is evident that the concentration of total alkaloids was maximum in distilled water extract (35.85mg/gm), followed by methanol extract (17.65mg/gm), ethanol extract (15.75mg/gm), petroleum ether extract (14.75mg/gm), acetone extract (12.45mg/gm) and benzene extract (11.75mg/gm). The concentration of total flavonoids was maximum in ethanol and methanol extracts (42.45mg/gm and 42.65mg/gm respectively),

followed by distilled water extract (27.87mg/gm), benzene extract (13.35mg/gm), petroleum ether extract (14.65mg/gm) and acetone extract (13.55mg/gm). The amount of total phenol was maximum in ethanol and methanol extracts (15.35 and 15.45mg/gm respectively), followed by distilled water extract (12.33mg/gm), benzene and petroleum ether extracts (9.85 and 11.67mg/gm respectively) and acetone extract (10.65mg/gm). Saponin concentration was maximum in ethanol (16.75mg/gm), benzene extract (15.25mg/gm) and distilled water extract (15.35mg/gm). Acetone and petroleum ether extracts contained relatively least amount of saponins (10.55 and 11.45mg/gm respectively). The total tannin concentration was maximum in ethanol extract (12.25mg/gm), followed by petroleum ether, acetone and benzene extracts (11.55, 10.35 and 10.35 mg/gm respectively. Methanol and distilled water extracts contained relatively low amount of total tannins, 9.25 mg/gm and 9.45 mg/gm respectively (Table- 3; Fig- 2).

IV. Discussion

The leaf extracts of *centella asiatica* showed the presence of terpenoids, steroids and phytosterols, tannins, alkaloids, glycosides, saponins, reducing sugars, phenols and flavonoids. The extraction of various phytochemicals was seen to be more effectively done in polar solvents (ethanol, methanol and distilled water) than the non polar (Acetone, petroleum ether, benzene) solvents. Especially, ethanolic, methanolic and distilled water leaf extracts showed presence of most of the tested phytochemicals. Hence, it can be reported that alcoholic extract was the best one for extracting the active principle than others. Flavonoids are water-soluble polyphenolic compounds which are extremely common and widespread in the plant kingdom as their glycosides. The flavonoids are known to act through scavenging or chelating process. The present findings gain support from the work of Arpita Roy et al., (2018) [84] who have found a more or less similar phytochemicals qualitatively and quantitatively in Centella asiatica. They observed similar phytochemicals in extracts of whole plant, shoot culture, callus culture and suspension culture. A more or less similar results was also observed by Rupa et al., (2017) [85]. According to reports, Centella asiatica extracts by ultrasonic assisted extraction showed Total Phenolic Content of 1350 mg GAE/100 g Dry Weight and Total Flavonoid Content, 599 mg QE/100 g Dry weight (Nithyanadam et al., (2014) [86]. The polyphenols in 100% ethanol extract was 21.1 ± 0.1 Pyrogallol Equivalent and flavanoid is 9.3 ± 0.3 Quercetin Equivalent (Rahman et al., (2013) [87]. The polyphenols in Centella asiatica was found to be 150 mg tannic acid/100 g for C. asiatica (Gupta et al., (2013) [88].

V. Conclusions

The results obtained in the present investigation are encouraging and can be used as reference data for the standardization of leaves of *Centella asiatica* and the formulations containing these plant leaves as a main ingredient. The evaluation of the various proximate parameters for the leaves of *Centella asiatica* has given a clear idea about the specific characteristics of these crude drugs under examination, in their powder form. The preliminary screening tests may be useful in the detection of the bioactive principles and subsequently may lead to the drug discovery and development. Such screening experiments form a primary platform for further phytochemicals and pharmacological studies that may open the possibility of finding new clinically effective compounds. Thus, the present study has authenticated the usefulness of useful drugs due to their rich contents of phytochemicals. The data obtained in the present study is expected to serve as valuable tool for identification, authentication and detection of adulterants, standardization and quality control of the drugs. Hence it can be concluded that the results of the present study have given qualitative and quantitative information about the purity standards of the leaves of *Centella asiatica*.

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References

- [1]. Shanghai PR. (1977): Jiangsu new medical college. Dictionary of Chinese Materia Medica. China: Shanghai Scientific and Technical Publishing House; 1977. p. 1874.
- [2]. Yu QL, Duan HQ, Takaishi Y, Gao WY. (2006): A novel triterpene from Centella asiatica. Molecules, 11(9):661-5.
- [3]. Jamil SS, Nizami Q, Salam M. (2007): Centella asiatica (Linn.) Urban-a review. Nat Prod Radiance, 6(2):158-70.
- [4]. Diwan PV, Karwande I, Singh AK. (1991): Anti-anxiety profile of manduk parni (*Centella asiatica*) in animals. *Fitoterapia*, 62:253-7.
 [5] M. Nir, H. Wir, J. J. St. H. (1994). Phys. Rev. B 14, 167 (1994).
- [5]. Wagner H, Nörr H, Winterhoff H. (1994): Plant adaptogens. *Phytomedicine*, 1(1):63-76. Mishra LC, editor. Scientific Basis for Ayurvedic Therapies. New York: CRC Press; 2003.
- [6]. Mishra LC, editor. (2003): Scientific Basis for Ayurvedic Therapies. New York: CRC Press

- [7]. Mishra LC, editor. Scientific Basis for Ayurvedic Therapies. New York: CRC Press; 2003.
- [8]. Mishra LC, editor. Scientific Basis for Ayurvedic Therapies. New York: CRC Press; 2003.
- [9]. Mishra LC, editor. Scientific Basis for Ayurvedic Therapies. New York: CRC Press; 2003.
- [10]. Bown D. (1995): Encyclopaedia of Herbs and their Uses. London: Dorling Kindersley; 1995.p. 361-5.
- [11]. Chevallier A. (1996): The Encyclopedia of Medicinal Plants. London: Dorling Kindersley; 1996.p. 257.
- [12]. Chopra RN, Nayar SL, Chopra IC.(1986): Glossary of Indian Medicinal Plants (Including the Supplement). New Delhi: Council of Scientific and Industrial Research; 1986.p. 51-83.
- [13]. Pandey NK, Tewari KC, Tewari RN, Joshi GC, Pande VN and Pandey G., (1993): Medicinal plants of Kumaon Himalaya, strategies for conservation, In: Dhar U (ed) Himalaya Biodiversity Conservation Strategies, *Himavikas Publication*, Nanital, No 3, 1993, 293-302.
- [14]. Singh HG., (1989): Himalayan herbs and drugs, importance and extinction threat, J. Sci Res. Plants Med, 10: 47-52.
- [15]. Sharma BL and Kumar A., (1998): Biodiversity of medicinal plants of Triyugi Narain (Garhwal Himalaya) and their conservation, National conference on recent trends in spices and medicinal plant research, A-78 (2-4 April). Calcutta, WB. India,
- [16]. Devkota Anjana and Jha Kumar Pramod, (2009): Variation in growth of *Centella asiatica* along different soil composition, *Botany Research International*, **2**(1): 55-60.
- [17]. Subban Ravi, Veerakumar A., Manimaran R., Hashim K.M., Balachandra Indira, (2008): Two new flavonoids from Centella asiatica (Linn.), J Nat Med., 62:369-373.
- [18]. Roy DC, Barman SK, Shaik MM. (2013): Current updates on *Centella asiatica*: Phytochemistry, pharmacology and traditional uses. *Med Plant Res*, 3(4):70-7.
- [19]. James JT, Dubery IA. (2009): Pentacyclic triterpenoids from the medicinal herb, *Centella asiatica* (L.) Urban. *Molecules*, **14(10)**:3922-41.
- [20]. Siddiqui BS, Aslam H, Ali ST, Khan S, Begum S. (2007): Chemical constituents of *Centella asiatica*. J Asian Nat Prod Res, 9(3-5):407-14.
- [21]. Inamdar PK, Yeole RD, Ghogare AB, De Souza NJ. (1996): Determination of biologically active constituents in *Centella asiatica*. J Chromatogr A, 742(1-2):127-30.
- [22]. Randriamampionona D, Diallo B, Rakotoniriana F, Rabemanantso C, Cheuk K, Corbisier AM, et al. (2007): Comparative analysis of active constituents in *Centella asiatica* samples from Madagascar: Application for ex situ conservation and clonal propagation. *Fitoterapia*, **78**(7-8):482-9.
- [23]. Yu QL, Duan HQ, Gao WY, Takaishi Y. (2007): A new triterpene and a saponin from *Centella asiatica*. *Chin Chem Lett*, **18**(1):62-4.
- [24]. Singh P, Singh JS. (2002): Recruitment and competitive interaction between ramets and seedlings in a perennial medicinal herb, *Centella asiatica. Basic Appl Ecol*, **3:**65-76.
- [25]. PDR for herbal medicine. (1999): 1st ed. Montvale, NJ: Medical Economics Co; 1999. p.729.
- [26]. Hagemann RC, Burnham TH, Granick B, Neubauer D. (1996): Gotu Kola, In, The Lawrence Review of Natural Products: facts and comparisons. St. Louis, MO, Facts and Comparisons Division, *J. B. Lippincott Co.*, p. 41-2.
- [27]. Baily E. (1945): Treatment of leprosy. Nature 1945;155:601.
- [28]. Wang L, Xu J, Zhao C, Zhao L, Feng B. (2013): Antiproliferative, cell-cycle dysregulation effects of novel asiatic acid derivatives on human non-small cell lung cancer cells. *Chem Pharm Bull* (Tokyo), **61**(10):1015-23.
- [29]. Pittella F, Dutra RC, Junior DD, Lopes MT, Barbosa NR. (2009): Antioxidant and cytotoxic activities of *Centella asiatica* (L) Urb. Int J Mol Sci, 10(10):3713-21.
- [30]. Babykutty S, Padikkala J, Sathiadevan P, Vijayakurup V, Azis T, Srinivas P, *et al.* (2009): Apoptosis induction of *Centella asiatica* on human breast cancer cells. *Afr J Tradit Complement Altern Med*, **6**(1):9-16.
- [31]. Hussin F, Eshkoor SA, Rahmat A, Othman F, Akim A. (2014): The centella asiatica juice effects on DNA damage, apoptosis and gene expression in hepatocellular carcinoma (HCC). BMC *Complement Altern Med*, **14**:32.
- [32]. Wu T, Geng J, Guo W, Gao J, Zhu X. (2017): Asiatic acid inhibits lung cancer cell growth *in vitro* and *in vivo* by destroying mitochondria. *Acta Pharm Sin B*, **7**(1):65-72.
- [33]. Park BC, Bosire KO, Lee ES, Lee YS, Kim JA. (2005): Asiatic acid induces apoptosis in SK-MEL-2 human melanoma cells. *Cancer Lett*, 218(1):81-90.
- [34]. Zhang J, Ai L, Lv T, Jiang X, Liu F. (2013): Asiatic acid, a triterpene, inhibits cell proliferation through regulating the expression of focal adhesionkinase in multiple myeloma cells. *Oncol Lett*, **6(6)**:1762-6.
- [35]. Kwon KJ, Bae S, Kim K, An IS, Ahn KJ, An S, et al. (2014): Asiaticoside, a component of Centella asiatica, inhibits melanogenesis in B16F10 mouse melanoma. Mol Med Rep, 10(1):503-7.
- [36]. Zaidan MR, Noor Rain A, Badrul AR, Adlin A, Norazah A, Zakiah I. (2005): In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. Trop Biomed, 22(2):165-70.
- [37]. Oyedeji OA, Afolayan AJ. (2005): Chemical composition and antibacterial activity of the essential oil of *Centella asiatica*. Growing in South Africa. *Pharma Biol*, 43(3):249-52.
- [38]. Pitinidhipat N. (2015): Antibacterial activity of *Chrysanthemum indicum, Centella asiatica* and *Andrographis paniculata* against *Bacillus cereus* and *Listeria monocytogenes* under osmotic stress. AUJT, **15(4)**:239-45.
- [39]. Sekar T, Ayyanar M, Pillai YJ. (2011): Phytochemical screening and antibacterial activity of leaf and callus extracts of *Centella* asiatica. Bangladesh J Pharmacol, 6(1):55-60.
- [40]. Dash BK, Faruquee HM, Biswas SK, Alam MK, Sisir SM, Prodhan UK. (2011): Antibacterial and antifungal activities of several extracts of *Centella asiatica* L. against some human pathogenic microbes. *Life Sci Med Res*, 2011:1-5.
- [41]. Dhiman R, Aggarwal N, Aneja KR, Kaur M. (2016): In vitro antimicrobial activity of spices and medicinal herbs against selected microbes associated with juices. Int J Microbiol, 9015802.
- [42]. Idris NA, Nadzir MM. (2017): Antimicrobial activity of *Centella asiatica* on *Aspergillus niger* and *Bacillus subtilis*. *Chem Eng Trans*, **56**:1381-6.
- [43]. Sultan RA, Mahmood SB, Azhar I, Ahmed SW, Mahmood ZA. (2014): Biological activities assessment of *Centella asiatica* (Linn.). J Herbs Spices Med Plants, **20**(3):319-27.
- [44]. Prakash V. (2017): Terpenoids as source of anti-inflammatory compounds. Asian J Pharm Clin Res, 10(3):68-76.
- [45]. Somchit MN, Sulaiman MR, Zuraini A, Samsuddin L, Somchit N, Israf DA, *et al.* (2004): Antinociceptive and antiinflammatory effects of *Centella asiatica*. *Indian J Pharmacol*, **36**(6):377.
- [46]. George M, Joseph L, Ramaswamy. (2009): Anti-allergic, anti-pruritic, and anti-inflammatory activities of *Centella asiatica* extracts. *Afr J Tradit Complement Altern Med*, 6(4):554-9.

- [47]. Huang SS, Chiu CS, Chen HJ, Hou WC, Sheu MJ, Lin Y, *et al.* (2011): Antinociceptive activities and the mechanisms of antiinflammation of asiatic acid in mice. *Evid Based Complement Alternat Med*, 2011:895857.
- [48]. Saha S, Guria T, Singha T, Maity TK. (2013): Evaluation of analgesic and anti-inflammatory activity of chloroform and methanol extracts of *Centella asiatica* Linn. ISRN *Pharmacol*, 2013:789613.
- [49]. Orhan IE. (2012): Centella asiatica (L.) Urban: From traditional medicine to modern medicine with neuroprotective potential. Evid Based Complement Altern Med 2012:946259.
- [50]. Defillipo PP, Raposo AH, Fedoce AG, Ferreira AS, Polonini HC, Gattaz WF, *et al.* (2012): Inhibition of cPLA2 and sPLA2 activities in primary cultures of rat cortical neurons by *Centella asiatica* water extract. *Nat Prod Commun*, **7**(7):841-3.
- [51]. Nasir MN, Habsah M, Zamzuri I, Rammes G, Hasnan J, Abdullah J. (2011): Effects of asiatic acid on passive and active avoidance task in male Spraque Dawley rats. *J Ethnopharmacol*, **134**(2):203-9.
- [52]. Thomas TN, Thomas PM. (2015): Patent Application No. 14/848,372. U.S; 2015.
- [53]. Prakash A, Kumar A. (2013): Mitoprotective effect of *Centella asiatica* against aluminum-induced neurotoxicity in rats: Possible relevance to its anti-oxidant and anti-apoptosis mechanism. *Neurol Sci*, **34**(8):1403-9.
- [54]. Gopi M, Arambakkam Janardhanam V. (2017): Asiaticoside: Attenuation of rotenone induced oxidative burden in a rat model of hemiparkinsonism by maintaining the phosphoinositide-mediated synaptic integrity. *Pharmacol Biochem Behav*, **155:**1-15.
- [55]. Hussin M, Abdul-Hamid A, Mohamad S, Saari N, Ismail M, Bejo MH.(2007): Protective effect of *Centella asiatica* extract and powder on oxidative stress in rats. *Food Chem*, 100(2):535-41.
- [56]. Raza SA, Adnan A, Qureshi F. (2009): Comparison of antioxidant activity of essential oil of *Centella asiatica* and Butylatedhydroxyanisole in sunflower oil at ambient conditions. *Biharean Biol*, **3**(1):71-5.
- [57]. Chandrika UG, Kumara PA. (2015): Gotu Kola (*Centella asiatica*): Nutritional properties and plausible health benefits. *Adv Food Nutr Res*, **76**:125-57.
- [58]. Jayashree G, Kurup Muraleedhara GK, Sudarslal S, Jacob VB. (2003): Anti-oxidant activity of *Centella asiatica* on lymphomabearing mice. *Fitoterapia*, **74**(**5**):431-4.
- [59]. Anand T, Mahadeva N, Phani KG, Farhath K. (2010): Antioxidant and DNA damage preventive properties of *Centella asiatica* (L) *Urb. Pharm J*, 2(17):53-8.
- [60]. Shetty BS, Udupa SL, Udupa AL, Somayaji SN. (2006): Effect of *Centella asiatica* L (Umbelliferae) on normal and dexamethasone-suppressed wound healing in Wistar Albino rats. *Int J Low Extrem Wounds*, **5(3)**:137-43.
- [61]. Somboonwong J, Kankaisre M, Tantisira B, Tantisira MH. (2012): Wound healing activities of different extracts of *Centella* asiatica in incision and burn wound models: An experimental animal study. *BMC Complement Altern Med*, **12**:103.
- [62]. Yao CH, Yeh JY, Chen YS, Li MH, Huang CH. (2017): Wound-healing effect of electrospun gelatin nanofibres containing *Centella* asiatica extract in a rat model. J Tissue Eng Regen Med, **11**(3):905-15.
- [63]. Chen Y, Han T, Qin L, Rui Y, Zheng H. (2003): Effect of total triterpenes from *Centella asiatica* on the depression behavior and concentration of amino acid in forced swimming mice. Zhong Yao Cai **26**(**12**):870-3.
- [64]. Kalshetty P, Aswar U, Bodhankar S, Sinnathambi A, Mohan V, Thakurdesai P. (2012): Antidepressant effects of standardized extract of *Centella asiatica* L in olfactory bulbectomy model. *Biomed Aging Pathol*, **2**(2):48-53.
- [65]. Ceremuga TE, Valdivieso D, Kenner C, Lucia A, Lathrop K, Stailey O, Taylor A. (2015): Evaluation of the anxiolytic and antidepressant effects of asiatic acid, a compound from Gotu kola or *Centella asiatica*, in the male Sprague Dawley rat. *AANA J*, **83(2):**91-8.
- [66]. Rahman MM, Sayeed MS, Haque MA, Hassan MM, Islam SA. (2012) : Phytochemical screening, antioxidant, anti-Alzheimer and anti-diabetic activities of *Centella asiatica. J Nat Prod Plant Resour*, **2(4)**:504-11.
- [67]. Gayathri V, Lekshmi P, Padmanabhan RN. (2011): Anti-diabetes activity of ethanol extract of *Centella asiatica* (L.) urban (whole plant) in Streptozotocin-induced diabetic rats, isolation of an active fraction and toxicity evaluation of the extract. *Int J Med Aromat Plants*, **1(3)**:278-6.
- [68]. Supkamonseni N, Thinkratok A, Meksuriyen D, Srisawat R. (2014): Hypolipidemic and hypoglycemic effects of *Centella asiatica* (L.) extract *in vitro* and *in vivo*. *Indian J Exp Biol*, **52(10)**:965-71.
- [69]. Haque S, Naznine T, Ali M, Azad TT, Morshed T, Afsana NA, et al. (2013): Antihyperglycemic activities of leaves of Brassica oleracea, Centella asiatica and Zizyphus mauritiana: Evaluation through oral glucose tolerance tests. Adv Nat Appl Sci, 7(5):519-26.
- [70]. Kabir AU, Samad MB, D'Costa NM, Akhter F, Ahmed A, Hannan JM. (2014): Anti-hyperglycemic activity of *Centella asiatica* is partly mediated by carbohydrase inhibition and glucose-fiber binding. *BMC Complement Altern Med*, **14**:31.
- [71]. Maulidiani, Abas F, Khatib A, Perumal V, Suppaiah V, Ismail A, *et al.* (2016): Metabolic alteration in obese diabetes rats upon treatment with *Centella asiatica* extract. *J Ethnopharmacol*, **180**:60-9.
- [72]. Wang X, Lu Q, Yu DS, Chen YP, Shang J, Zhang LY, *et al.* (2015): Asiatic acid mitigates hyperglycemia and reduces islet fibrosis in Goto-Kakizaki rat, a spontaneous Type 2 diabetic animal model. *Chin J Nat Med*, **13**(7):529-34.
- [73]. Liu J, He T, Lu Q, Shang J, Sun H, Zhang L. (2010): Asiatic acid preserves beta cell mass and mitigates hyperglycemia in streptozocin-induced diabetic rats. *Diabetes Metab Res Rev*, **26(6):**448-54.
- [74]. Chaisawang P, Sirichoat A, Chaijaroonkhanarak W, Pannangrong W, Sripanidkulchai B, Wigmore P, *et al.* (2017): Asiatic acid protects against cognitive deficits and reductions in cell proliferation and survival in the rat hippocampus caused by 5-fluorouracil chemotherapy. *PLoS One*, **12**(7):e0180650.
- [75]. Gray NE, Zweig JA, Murchison C, Caruso M, Matthews DG, Kawamoto C, et al. (2017): Centella asiatica attenuates Aß-induced neurodegenerative spine loss and dendritic simplification. Neurosci Lett, 646:24-9.
- [76]. Farhana KM, Malueka RG, Wibowo S, Gofir A. (2016): Effectiveness of Gotu Kola extract 750?mg and 1000?mg compared with folic acid 3?mg in improving vascular cognitive impairment after stroke. *Evid Based Complement Alternat Med*, **2016**:2795915.
- [77]. Umka Welbat J, Sirichoat A, Chaijaroonkhanarak W, Prachaney P, Pannangrong W, Pakdeechote P, et al. (2016): Asiatic acid prevents the deleterious effects of valproic acid on cognition and hippocampal cell proliferation and survival. Nutrients, 8(5). pii: E303.
- [78].]. Gray NE, Harris CJ, Quinn JF, Soumyanath A. (2016): *Centella asiatica* modulates antioxidant and mitochondrial pathways and improves cognitive function in mice. *J Ethnopharmacol*, **180**:78-6.
- [79]. Gray NE, Sampath H, Zweig JA, Quinn JF, Soumyanath A. (2015): *Centella asiatica* attenuates amyloid-ß-induced oxidative stress and mitochondrial dysfunction. J Alzheimers Dis, 45(3):933-46.
- [80]. Oyenihi AB, Chegou NN, Oguntibeju OO, Masola B. (2017): *Centella asiatica* enhances hepatic antioxidant status and regulates hepatic inflammatory cytokines in Type 2 diabetic rats. Pharm Biol, 55(1):1671-8.
- [81]. Choi MJ, Zheng HM, Kim JM, Lee KW, Park YH, Lee DH. (2016): Protective effects of *Centella asiatica* leaf extract on dimethylnitrosamine-induced liver injury in rats. *Mol Med Rep* **14**(**5**):4521-8.

- [82]. Ghosh K, Indra N, Jagadeesan G. (2017): The ameliorating effect of *Centella asiatica* ethanolic extract on albino rats treated with isoniazid. *J Basic Clin Physiol Pharmacol*, **28**(1):67-7.
- [83]. Duggina P, Kalla CM, Varikasuvu SR, Bukke S, Tartte V. (2015): Protective effect of centella triterpene saponins against cyclophosphamide-induced immune and hepatic system dysfunction in rats: Its possible mechanisms of action. *J Physiol Biochem*, **71(3)**:435-4.
- [84]. Tang LX, He RH, Yang G, Tan JJ, Zhou L, Meng XM, *et al.* (2012): Asiatic acid inhibits liver fibrosis by blocking TGFbeta/Smad signaling *in vivo* and *in vitro*. *PLoS One*, **7**(2):e31350.
- [85]. Rajalingam D, Varadharajan R, Palani S. (2016): Evaluation of hepatoprotective and antioxidant effect of *Combretum albidum* G. don against CCl4 induced hepatotoxicity in rats. *Int J Pharm Pharm Sci*, **8**(9):218-23.
- [86]. Harborne JB. (1973): Phytochemicals Methods. Chapman and Hall Ltd, London, 1973, 49-188.
- [87]. Arpita Roy M, Krishnan L and Bharadvaja N (2018) Qualitative and Quantitative Phytochemical Analysis of Centella asiatica. Nat Prod Chem Res 6: 323. doi:10.4172/2329-6836.1000323
- [88]. Rupa D, Bhattacharya Komal M. Parmar, Prakash, R. Itankar, Satyendra, K. Prasad (2017): Phytochemical and pharmacological evaluation of organic and non-organic cultivated nutritional *Centella asiatica* collected after different time intervals of harvesting, *South African Journal of Botany*, **112**: 237-245
- [89]. Nithyanandam R, Shapheri MR, Nassir M (2014) Antioxidant potential of Malaysian herb Centella asiatica, 3rd international conference on environment. *Chem Bio* **78:** 1-7.
- [90]. Rahman M, Hossain S, Rahaman A, Fatima N, Nahar T, et al. (2013): Antioxidant activity of Centella asiatica (Linn.) Urban: Impact of extraction solvent polarity. J Pharmaco Phytochem 1: 27-32.
- [91]. Gupta S, Gowri BS, Lakshmi AJ, Prakash J (2013) Retention of nutrients in green leafy vegetables on dehydration. J Food Sci Technol **50**: 918-925.

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